



FastQC website:

http://www.bioinformatics.babraham.ac.uk

FastQC report documentation:

http://www.bioinformatics.babraham.ac.uk/projects/fastqc/Help/3%20Analysis%20Modules/

Good Illumina dataset:

http://www.bioinformatics.babraham.ac.uk/projects/fastqc/good_sequence_short_fastqc/fastqc_report.html

Bad Illumina dataset:

http://www.bioinformatics.babraham.ac.uk/projects/fastqc/bad_sequence_fastqc/fastqc_report.html

Real Yeast ChIP-seq dataset:

http://web.corral.tacc.utexas.edu/BiolTeam/yeast_stuff/Sample_Yeast_L005_R1.cat_fastqc/fastqc_report.html

Most useful reports

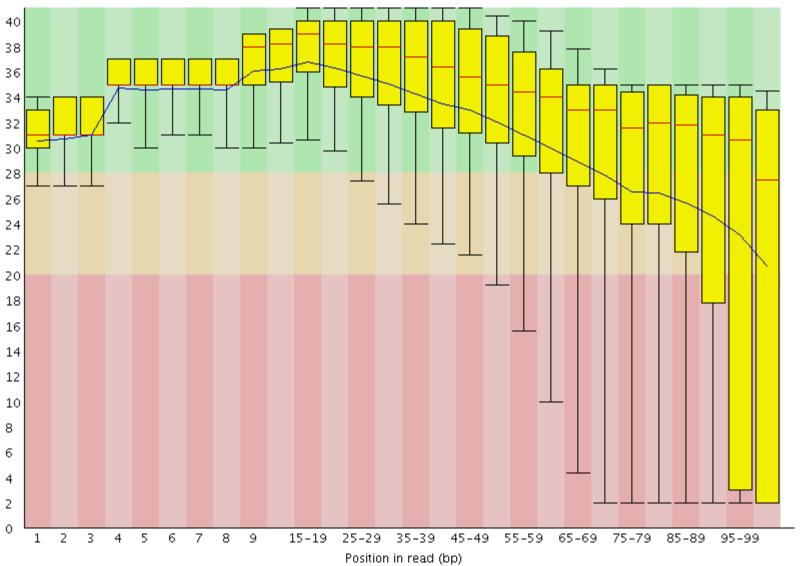


- Should I trim low quality bases?
 - Per-base sequence quality Report
 - based on all sequences
- Do I need to remove adapter sequences?
 - Overrepresented sequences Report
 - based on 1st 200,000 sequences
- How complex is my library?
 - Sequence duplication levels Report
 - estimate based on 1st 200,000 sequences

Per-base sequence quality











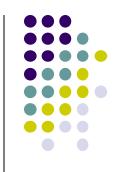
Sequence	Count	Percentage	Possible Source
AGATCGGAAGACCACGTCTGAACTCCAGTCACCTCAGAATCTCGTATG	60030	5.01369306977828	TruSeq Adapter, Index 1 (97% over 37bp)
GATCGGAAGAGCACACGTCTGAACTCCAGTCACCTCAGAATCTCGTATGC	42955	3.5875926338884896	TruSeq Adapter, Index 1 (97% over 37bp)
CACACGTCTGAACTCCAGTCACCTCAGAATCTCGTATGCCGTCTTCTGCT	3574	0.29849973398946483	RNA PCR Primer, Index 40 (100% over 41bp)
CAGATCGGAAGAGCACACGTCTGAACTCCAGTCACCTCAGAATCTCGTAT	2519	0.2103863542024236	TruSeq Adapter, Index 1 (97% over 37bp)
GAGATCGGAAGAGCACACGTCTGAACTCCAGTCACCTCAGAATCTCGTAT	1251	0.10448325887543942	TruSeq Adapter, Index 1 (97% over 37bp)

Overrepresented Sequences



Sequence	Count	Percentage	Possible Source
AACGACTCTCGGCAACGGATATCTCGGCTCTCGCATCGATGAAGAACGTA	102020	1.0707851766890004	No Hit
${\tt AATTCTAGAGCTAATACGTGCAACAAACCCCGACTTATGGAAGGGACGCA}$	89437	0.9387160737848865	No Hit
${\tt AAAGGATTGGCTCTGAGGGCTGGGCTCGGGGGTCCCAGTTCCGAACCCGT}$	89427	0.9386111154260659	No Hit
${\tt TACCTGGTTGATCCTGCCAGTAGTCATATGCTTGTCTCAAAGATTAAGCC}$	87604	0.9194772066130483	No Hit
${\tt ATTGGCTCTGAGGGCTGGGCTCGGGGTCCCAGTTCCGAACCCGTCGGCT}$	65829	0.6909303802809273	No Hit
${\tt TCTAGAGCTAATACGTGCAACAAACCCCGACTTATGGAAGGGACGCATTT}$	65212	0.6844544495416888	No Hit
${\tt TAAAACGACTCTCGGCAACGGATATCTCGGCTCTCGCATCGATGAAGAAC}$	61582	0.646354565289767	No Hit
$\tt CTCGGATAACCGTAGTAATTCTAGAGCTAATACGTGCAACAAACCCCGAC$	59180	0.6211435675010296	No Hit
${\tt ATGGATCCGTAACTTCGGGAAAAGGATTGGCTCTGAGGGCTGGGCTCGGG}$	56982	0.598073720232235	No Hit
${\tt AAAACGACTCTCGGCAACGGATATCTCGGCTCTCGCATCGATGAAGAACG}$	54813	0.5753082522040206	No Hit
${\tt ATTCTAGAGCTAATACGTGCAACAAACCCCGACTTATGGAAGGGACGCAT}$	52688	0.5530046009546172	No Hit
${\tt GCGACCCCAGGTCAGGCGGGATTACCCGCTGAGTTTAAGCATATCAATAA}$	41363	0.4341392595901502	No Hit
$\tt CTAGAGCTAATACGTGCAACAAACCCCGACTTATGGAAGGGACGCATTTA$	40019	0.4200328561646452	No Hit
${\tt AGAACTCCGCAGTTAAGCGTGCTTGGGCGAGAGTAGTACTAGGATGGGTG}$	39753	0.4172409638200141	No Hit
ACTCGGATAACCGTAGTAATTCTAGAGCTAATACGTGCAACAAACCCCGA	38867	0.4079416532284981	No Hit
${\tt ACGACTCTCGGCAACGGATATCTCGGCTCTCGCATCGATGAAGAACGTAG}$	38438	0.40343893963508914	No Hit
${\tt ACTTCGGGAAAAGGATTGGCTCTGAGGGCTGGGCTCGGGGGTCCCAGTTC}$	37406	0.3926072370047907	No Hit
AGATCGGAAGAGCACACGTCTGAACTCCAGTCACTGACCAATCTCGTATG	34199	0.35894709133098535	TruSeq Adapter, Index 4 (100% over 49bp)
${\tt GAACCTTGGGATGGGTCGGCCGGTCCGCCTTTGGTGGTCATTGGTCGGCT}$	34099	0.3578975077427782	No Hit

Overrepresented Sequences



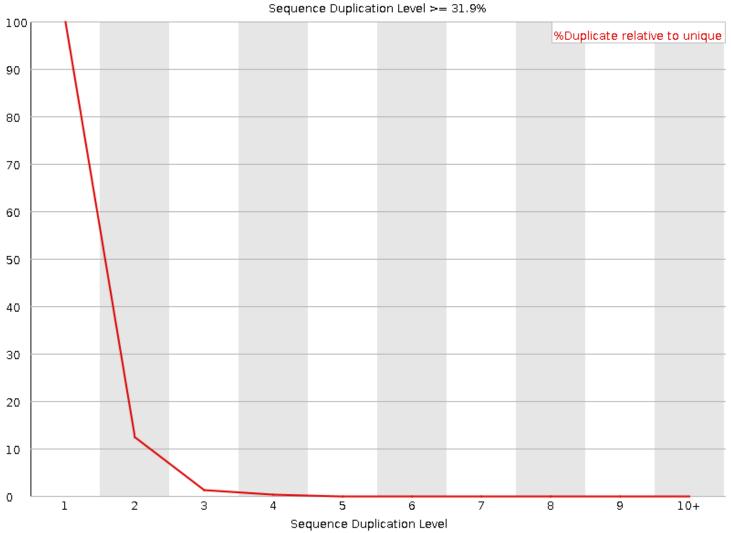
Sequence	Count	Percentage	Possible Source
GAAGGTCACGGCGAGACGAGCCGTTTATCATTACGATAGGTGTCAAGTGG	5632816	32.03026785752871	No Hit
${\tt TATTCTGGTGTCCTAGGCGTAGAGGAACAACACCAATCCATCC$	494014	2.8091456822607364	No Hit
${\tt TCAAACGAGGAAAGGCTTACGGTGGATACCTAGGCACCCAGAGACGAGGA}$	446641	2.539765344040083	No Hit
${\tt TAAAACGACTCTCGGCAACGGATATCTCGGCTCTCGCATCGATGAAGAAC}$	179252	1.0192929387357474	No Hit
${\tt GAAGGTCACGGCGAGACGAGCCGTTTATCATTACGATAGGGGTCAAGTGG}$	171681	0.9762414422996221	No Hit
${\tt AACGACTCTCGGCAACGGATATCTCGGCTCTCGCATCGATGAAGAACGTA}$	143415	0.8155105483274229	No Hit
${\tt AGAACATGAAACCGTAAGCTCCCAAGCAGTGGGAGGAGCCCTGGGCTCTG}$	111584	0.6345077504066322	No Hit
${\tt AAAACGACTCTCGGCAACGGATATCTCGGCTCTCGCATCGATGAAGAACG}$	111255	0.6326369351474214	No Hit
${\tt ATTACGATAGGTGTCAAGTGGAAGTGCAGTGATGTATGCAGCTGAGGCAT}$	73682	0.41898300890326096	No Hit
${\tt GAAGGTCACGGCGAGACGAGCCGTTTATCATTACGATAGGTGTCAAGGGG}$	71661	0.4074908580252516	No Hit
$\tt GGATGCGATCATACCAGCACTAATGCACCGGATCCCATCAGAACTCCGCA$	69548	0.3954755612388914	No Hit
${\tt ATATTCTGGTGTCCTAGGCGTAGAGGAACAACACCAATCCATCC$	54017	0.30716057099328803	No Hit

Duplication levels Pences Yeast ChIP-seq

for every 100 unique sequences there are

- ~12 sequences w/2 copies
- ~1-2 with 3 copies



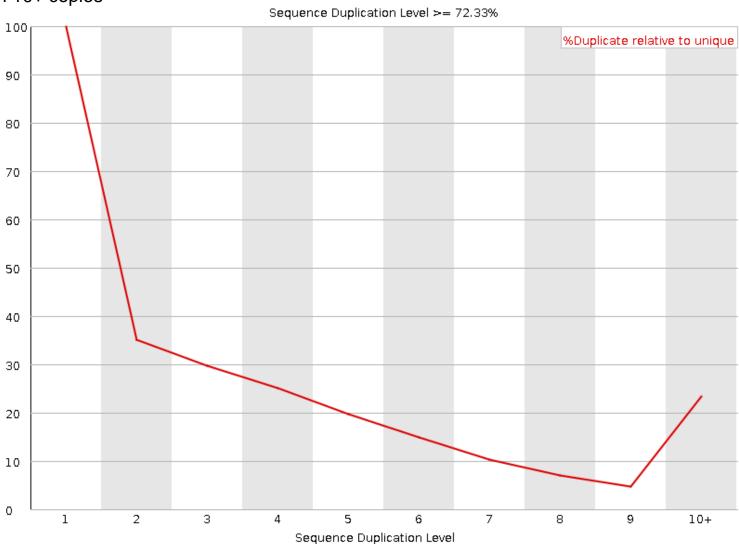


Duplication levels

Yeast ChIP-exo

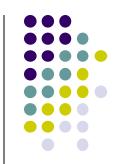
for every 100 unique sequences there are

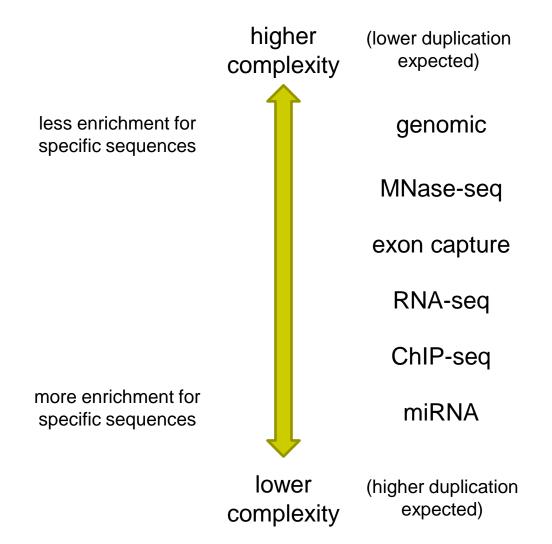
- ~35 sequences w/2 copies
- ~22 with 10+ copies





Library complexity is a function of experiment type (& sequencing depth)





Running FastQC



- Can run as interactive tool or command line
- Input:
 - fastq files (R1, R2 separately)
- Output:
 - directory with html & text reports
 - fastqc_report.html
 - fastqc_data.txt