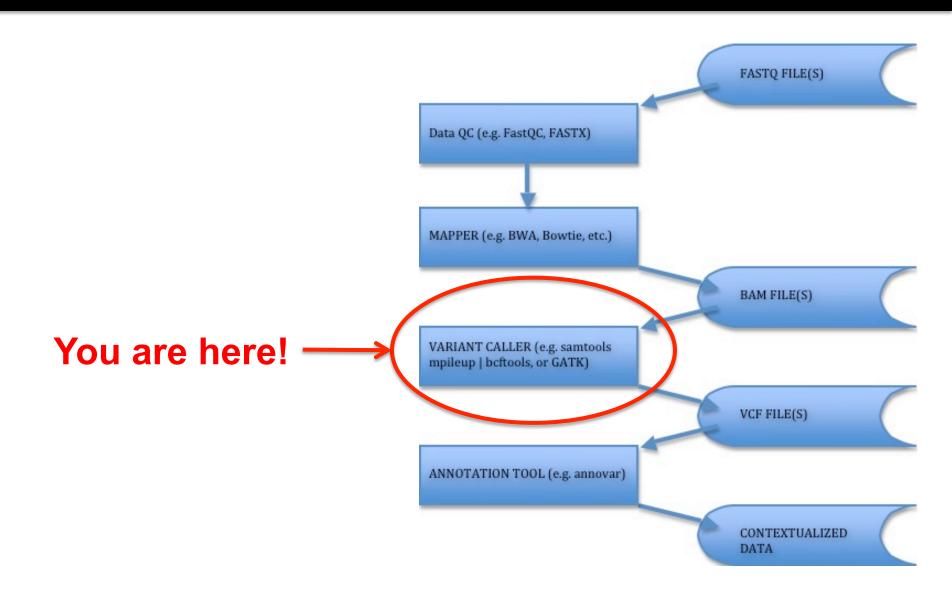
Advanced Variant Calling



Types of Genome Sequence Variants

Single Nucleotide Variants (SNVs) *

Single base changes, e.g., A→T.

2. Insertions-Deletions (Indels; DIPs) ★▶

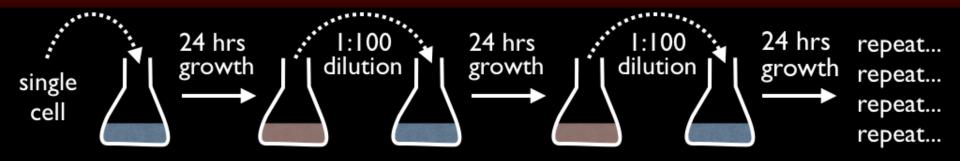
Consisting of one or a few bases, e.g., +ATGA, ΔT.

3. Structural Variants (SVs) ▶

 Everything else: large deletions, insertions, duplications, inversions, translocations, mobile element insertions, horizontal gene transfer

Different sequencing information and different algorithms are used to predict each kind of variant.

Long-term E. coli evolution experiment



- I2 independent populations evolved >20 yrs. Frozen "fossil record" has been archived.
- How many and what mutations?
- Compare rates of genomic change and fitness increase, monitor diversity in the population, understand molecular basis of adaptation.

Knowing what you don't know

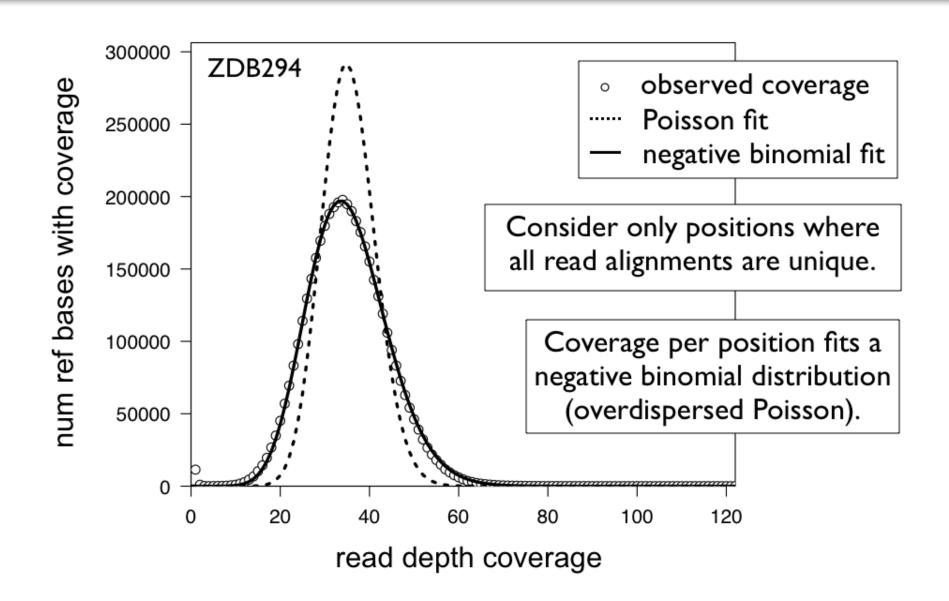
- 1. Theoretical limits: Read length and pair distance.
- 2. Practical limits: Base quality and coverage evenness.

	single-end	paired-end	mate-paired
IS insertions	*	*	*
duplications	*	*	*
inversions across IS	_	_	*
SNPs in repeats	_	_	*
insertion of new sec	– P	_	_

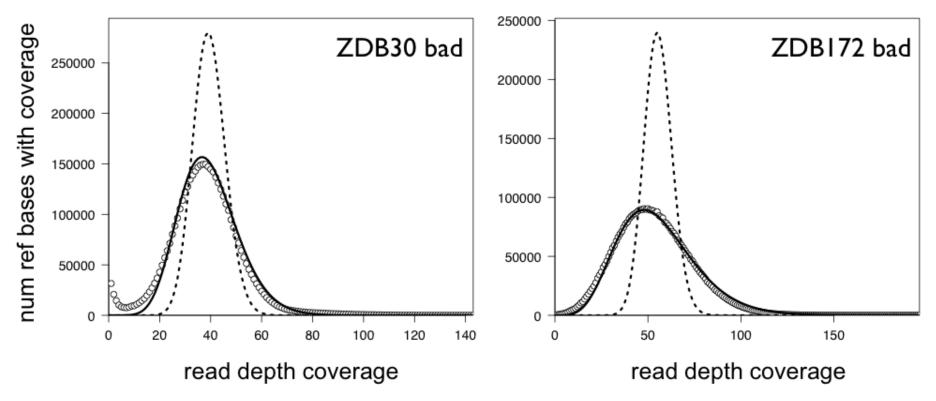
IS = bacterial mobile elements 0.8-1.5 kb in length.

Need standardized metrics to describe completeness of re-sequencing data on a per-base per-genome basis.

Typical Coverage Distribution



Problem Coverage Distributions



 Contamination with another sample? Large variance, missing coverage.

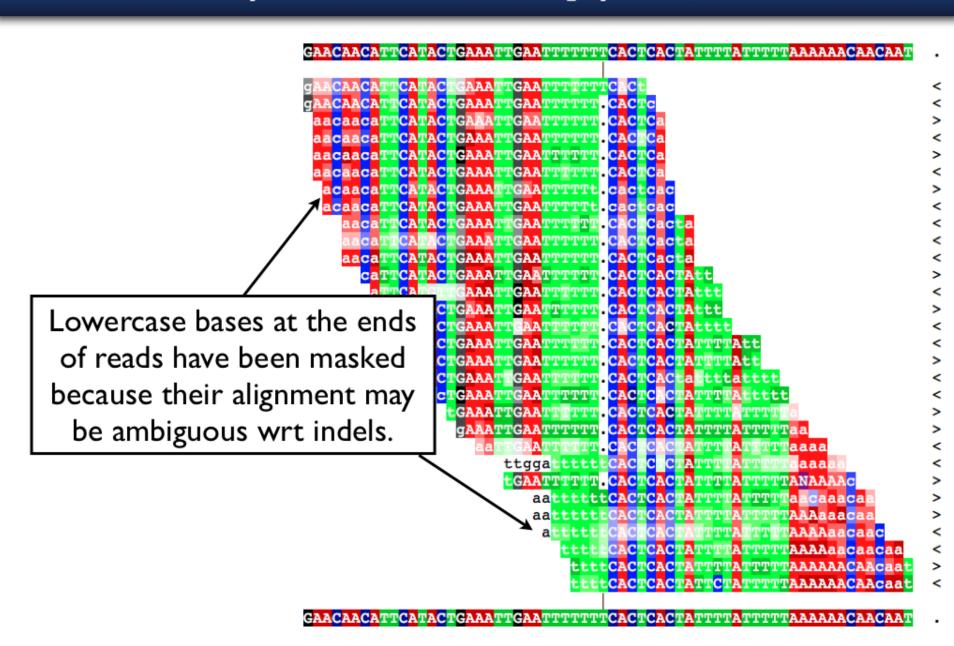
Both apparently from problems with library prep.

Identifying within-alignment indels

 Need to be careful in repetitive sequences and at the edges of short reads...

...where reads aligned from different directions can be ambiguously aligned.

...where reads from different directions that end in a simple sequence repeat may hide indels.



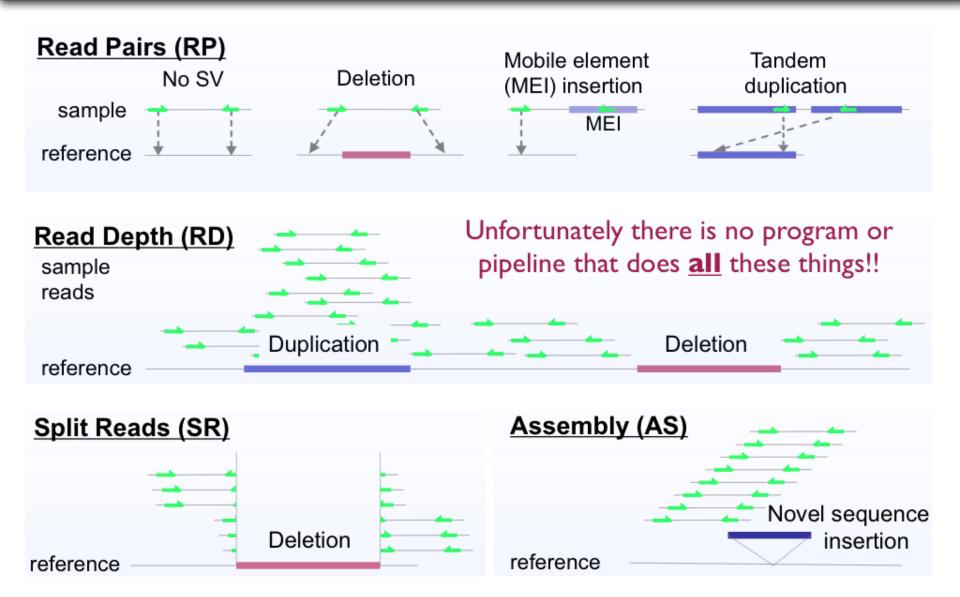
Pitfalls of the column mindset

```
GCGGGAGTGTCCGGGAATAA.T.T.AAAA.CGATGCACACAGGGTTTAGCGCGTA
                                                                     NC 001416-4/8118-8181
                                                                  < 4:34026/35-1 (MQ=21)
           CCGGGAATAA.T.TAAAAA.CGATGcaca
        rgrgcgggaataa. Tca. Aaaa. cgatgcacaccg
                                                                  < 3:4689/35-3 (MQ=255)
        GTCCGGGAATAA.T.TAAAAA.CGATGCACACAG
                                                                  > 5:37643/1-35 (MQ=255)
        GTCCGGGAATAA.T.TAAAAA.CGATGCACACAG
                                                                  > 1:34048/1-35 (MQ=255)
        rggggggaataa. Tca.aaaa. cgatgcacacc
                                                                  < 2:38949/35-3 (MQ=255)
        TGTGCGGGAATAA .TCA .AAAA .CGATGCACACCgg
TGTCCGGGAATAA .T.TAAAAA .CGATGCACACAgg
TGTCCGGGAATAA .T.TAAAAA .CGATGCACACAgg
                                                                  < 5:22681/35-1 (MQ=255)
                                                                  < 5:357/35-1 (MQ=255)
                                                                  < 2:12956/35-1 (MQ=255)
           GCGGGAATAA TCA AAAA CGATGCACAC
                                                                  > 3:23632/1-35 (MQ=255)
```

Requires local multiple sequence re-alignment to get it right!

Implemented in samtools mpileup and the Genome Alignment Toolkit (GATK).

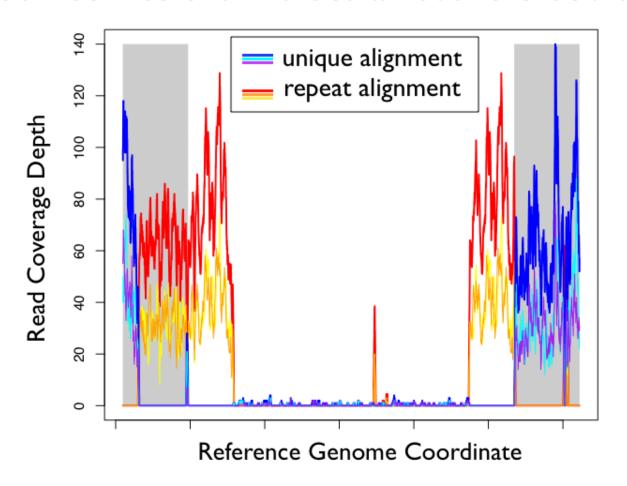
Predicting structural variants



Identifying large deletions

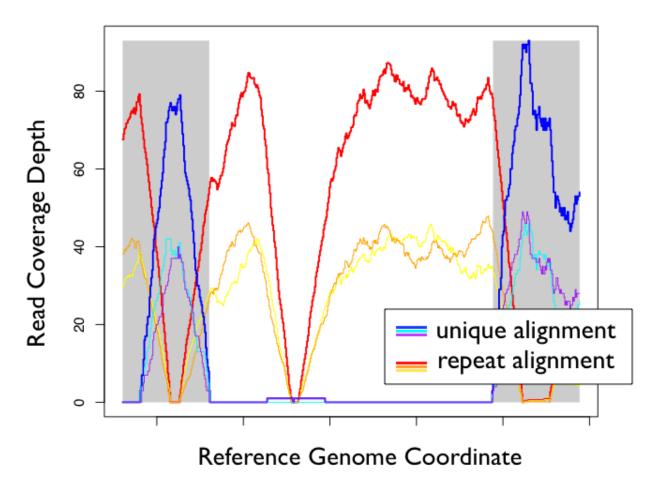
- 1. Seed deletions at positions with zero coverage.
- Propagate boundaries outward until reaching a readdepth threshold based on the overall distribution.
- Propagate through repeat regions, where a read aligns to multiple places in the genome.

Sometimes the molecular event is obvious...



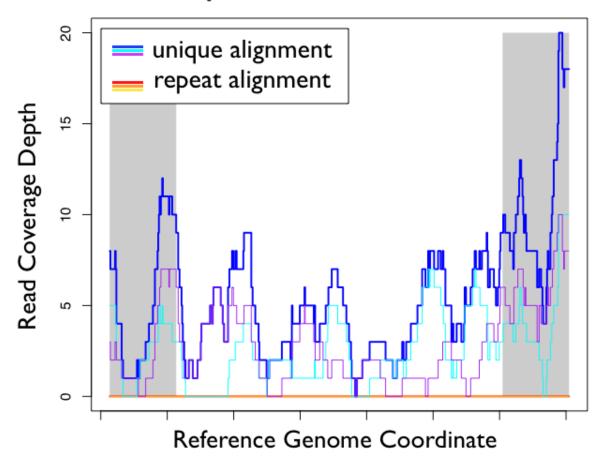
Recombination between nearby IS3 copies.

Sometimes the mutation is not obvious...



Gene conversion of 23S rRNA copy!!

 Sometimes overall low or biased coverage leads to false predictions of deletions.



Recognizable by sloped vs. steep edges.

Identifying new junctions

 Find "mosaic" reads that partially map to two locations in the genome (possibly with overlap).



- 2. Create consensus list of possible new junctions.
- 3. Re-align all reads to candidate junctions.



4. Predict a new junction if reads map better to it than to the reference across its whole length.

Example of a good junction

position	overlap	reads	gene	coords	product
1 =	0	36	-/thrL	/189	-/thr operon leader peptide
= 4629812			lasT/-	4629789/	predicted rRNA methyltransferase/-

```
REL606 1 1 REL606 4629812 0 0 /3-71
                                                                     30KR6AAXXLesnki set 1 2:3:53:1076:1729/1-36
              GCAGTCAGAATGAAAAGCTG
                                                                     30KR6AAXXLesnki set 1 2:3:3:1045:1537/1-36
                                                                     30KR6AAXXLesnki set 1 2:3:98:1256:1982/1-36
                                                                     30KR6AAXXLesnki set 1 2:3:69:59:1642/1-36
               AGTCAGAATGAAAAGCTGAAAAA
                                                                     30KR6AAXXLesnki set 1 2:3:52:1112:1970/1-36
                                                                     30KR6AAXXLesnki set 1 2:3:29:260:647/1-36
                                                                     30KR6AAXXLesnki set 1 2:3:45:197:1888/1-36
              GCAGTCAGAATGAAAAGCTGAAAAATAC
                                                                     30KR6AAXXLesnki set 1 2:3:38:829:160/1-36
                                                                     30KR6AAXXLesnki set 1 2:3:82:996:256/1-36
              GCAGTCAGAATGAAAAGCTGAAAAATACTTAC
                                                                     30KR6AAXXLesnki set 1 2:3:88:199:234/1-36
                                                                     30KR6AAXXLesnki set 1 2:3:31:1778:622/1-36
                                                                     30KR6AAXXLesnki set 1 2:3:21:1481:579/1-36
                GTCAGAATGAAAAGCTGAAAAATAC
                                                                     30KR6AAXXLesnki set 1 2:3:14:1273:59/1-36
                                                                     30KR6AAXXLesnki set 1 2:3:54:842:43/1-36
                GTCAGAATGAAAAGCTGAAAAATAC
                                                                     30KR6AAXXLesnki_set_1_2:3:82:844:525/1-36
                                                                     30KR6AAXXLesnki set 1 2:3:30:6:1419/1-36
                                                                     30KR6AAXXLesnki set 1 2:3:23:1578:360/1-36
                                                                     30KR6AAXXLesnki set 1 2:3:65:1765:1077/1-36
                                                                     30KR6AAXXLesnki set 1 2:3:62:1360:759/1-36
                                                                     30KR6AAXXLesnki set 1 2:3:65:842:32/1-36
                                                                     30KR6AAXXLesnki set 1 2:3:3:1093:1221/1-36
                                                                     30KR6AAXXLesnki set 1 2:3:13:204:1274/1-36
                                                                     30KR6AAXXLesnki set 1 2:3:8:699:65/1-36
                                                                     30KR6AAXXLesnki set 1 2:3:81:1575:760/1-36
                                                                     30KR6AAXXLesnki set 1 2:3:57:387:423/1-36
                                                                     30KR6AAXXLesnki set 1 2:3:19:601:1470/1-36
                                                                     30KR6AAXXLesnki set 1 2:3:71:503:526/1-36
                                                                     30KR6AAXXLesnki set 1 2:3:29:1139:1664/1-36
                                                                     30KR6AAXXLesnki set 1 2:3:71:505:527/1-36
                                                                     30KR6AAXXLesnki set 1 2:3:18:1079:1002/1-36
                                                                     30KR6AAXXLesnki set 1 2:3:6:1485:1308/1-36
                                                                     30KR6AAXXLesnki set 1 2:3:24:627:931/1-36
                                                                     30KR6AAXXLesnki set 1 2:3:92:145:1544/1-35
                                GAAAAA TACTTACTAAGGCG
                                                                     30KR6AAXXLesnki set 1 2:3:58:1720:1463/1-36
                                                                     30KR6AAXXLesnki set 1 2:3:86:300:312/1-36
                               TGAAAAATACTTACTAAGGCGTTTTTTAT
                                                                     30KR6AAXXLesnki set 1 2:3:41:1600:1707/1-36
REL606 1 1 REL606 4629812 0 0 /3-71
```

Example of a bad junction

Beware of reads ending in homopolymer runs!

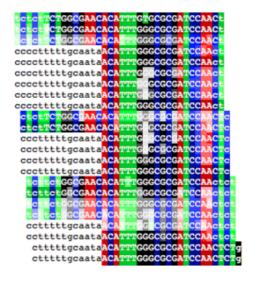
position	overlap	full / total reads	gene	coords	product
= 489705			ybbN	490447-489593	predicted thioredoxin domain-containing protein
3912264 =	0	7 / 14	ilvL/ilvG	3912221/3912359	ilvG operon leader peptide/acetolactate synthase II, valine insensitive, large subunit

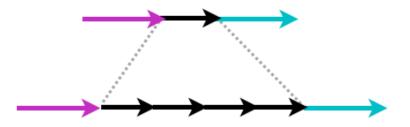
GAACGTTTTACGCGTCTGACCGTCTGCGGC.GG	REL606/489674-489705
GGGGG <mark>TTTTTTTTGACCTTA</mark>	REL606/3912264-3912283
CGCGTCTGACCGTCTGCGGC.GGCGCTTTTTTTTTC	30KR6AAXXLesnki_set_1_2:2:8:1611:1595
AGCCTTTTCTCGTTTC	30KR6AAXXLesnki_set_1_2:2:10:658:1121
TCTGACCGTCTGCGGC GGGGGTTTTTTTTTGAGGT	30KR6AAXXLesnki_set_1_2:2:13:1672:83
GCGTCTGACCGTCTGCGGC GGGGGGGTTTTTTTTTTTTT	30KR6AAXXLesnki_set_1_2:2:16:1338:1584
-AACGTTTTACGCGTCTGACCGTCTGCGGC	30KR6AAXXLesnki_set_1_2:2:29:1395:930
<mark>TTTTA</mark> CG <mark>CGTCTGACCGTCTGCGGC</mark> .GGGGGGGTGTT	30KR6AAXXLesnki_set_1_2:2:30:1685:1502
<mark>ACGTTTTACGCGTCTGACCGTCTGC</mark> GCGCCGT	30KR6AAXXLesnki_set_1_2:2:33:1415:263
<mark>TTTTACGCGTCTGACCGTCTGCGGC</mark> .GGGGGGTTT <mark>T</mark>	30KR6AAXXLesnki_set_1_2:2:37:666:557
-AACGTTTTACGCGTCTGACCGTCTGCGGC	30KR6AAXXLesnki_set_1_2:2:46:717:825
TTACTGGCTTTTGCACCCGGCGCGCTTTTTTTT	30KR6AAXXLesnki_set_1_2:2:59:1018:1338
<mark>CGTTTTA</mark> CGCG <mark>TCTGA</mark> CCG <mark>TCTGC</mark> GGC	30KR6AAXXLesnki_set_1_2:2:65:262:1990
GAACGTTTTACGCGTCTGACCGTCTGCGGC	30KR6AAXXLesnki_set_1_2:2:76:1050:1507
TACGCGTCTGACCGTCTGCGGC GGGGGGGTTTTTTT	30KR6AAXXLesnki_set_1_2:2:87:1336:724
TTTTACGCGTCTGACCGTCTGCGGC GGGGGGGTCTT	30KR6AAXXLesnki_set_1_2:2:99:618:1322
Base Quality Score Legend: Mcc < 22 ≤ M	$CC < 28 \le ATCC < 34 \le ATCC$

IS insertions create two new junctions...

		position	overlap	reads	gene	coords	product
		16989			IS150 (+)	+1443 (+3) bp	
3	?	2 16990 = 0 44	44	mokC/nhaA	I BUSU/I //IX /	regulatory protein for HokC, overlaps CDS of hokC/pH-dependent sodium/proton antiporter	
	2	= 3652533	3652533		IS150	3651091-3652533	repeat region
3	?	= 16992	0 41	41	mokC/nhaA		regulatory protein for HokC, overlaps CDS of hokC/pH-dependent sodium/proton antiporter
	?	2 3893554 =		IS150	3893554-3894996	repeat region	

Sometimes both new and old junctions exist...

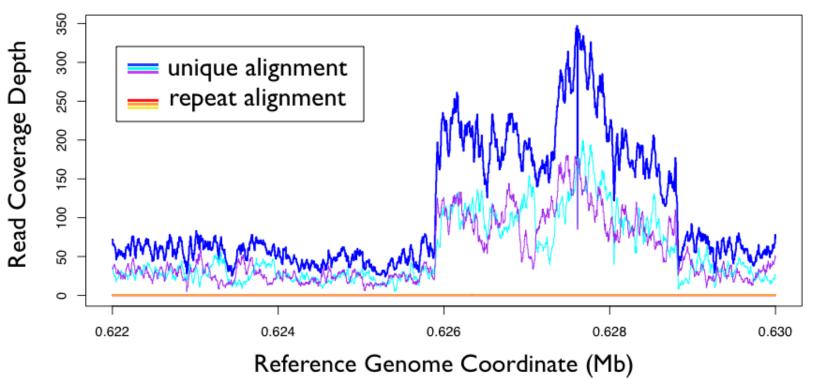




tandem head-to-tail duplications

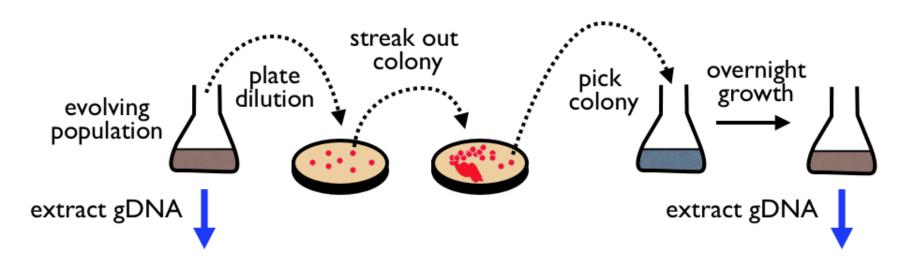
Identifying copy number variation

 Coverage is very noisy, but a fingerprint is (somewhat) consistent across runs.



 Tile into segments, train model on many genomes, look for deviation

Mixed population analysis



Every read could be from any individual.

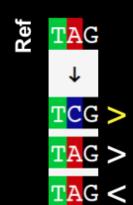
Frequencies of mutations competing in population.

No linkage information.

All reads are from a single clone.

Information about which mutations occur together.

Sequencing error or polymorphism?



TAG >

TCG >

TCG >

TAG <

TCG >

TCG >

TCG >

TAG >

Aligned

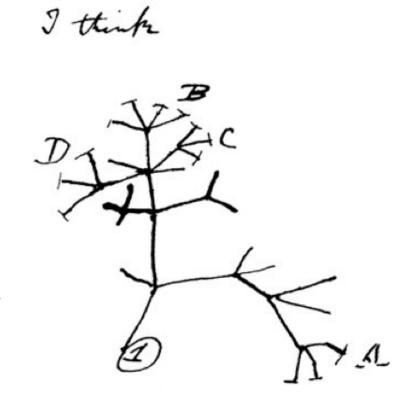
- Map reads to ancestor genome.
 Only consider single-base substitutions.
- Log-likelihood test for polymorphism:

$$D = -2 \ln \frac{\text{Pr (obs | no polymorphism, i.e. all error)}}{\text{Pr (obs | ML fraction new allele)}}$$

- Clone sequence data serves as a negative control (all errors, no polymorphisms).
- Filter out predictions with other biases: strand bias, systematically low quality scores

From evidence to mutations

- Genome sequencing data gives us <u>evidence</u> of changes: read alignments, missing coverage, new junctions, ...
- But we really want a list of biological <u>mutations</u> to study evolutionary history and molecular mechanisms.
- Complication: Later events may sometimes hide earlier events (e.g. SNV in region that is later deleted)



Genome Diffs

- To submit a changed genome sequence to GenBank you must currently re-submit the entire genome – even if it has only a one base difference.
- Mutational events are essentially genome differences.
 (In a Comp Sci sense of applying "patches" to files)
- Supplementary tables are not a sustainable, standardized, or re-usable way to report this data.
- An ideal genome analysis also reports what is not known, frequency information for mixed population samples, quality metrics, ...