ІТЕМ ТО СНЕСК	IMPORTANCE	CHECKLIST
EXPERIMENTAL DESIGN Definition of experimental and control groups	E	
Number within each group	<b>E</b> D	
Assay carried out by core lab or investigator's lab? Acknowledgement of authors' contributions	D	
SAMPLE Description	E	
Volume/mass of sample processed	D	
Microdissection or macrodissection Processing procedure	E E	
If frozen - how and how quickly?	E	
If fixed - with what, how quickly? Sample storage conditions and duration (especially for FFPE samples)	E E	
NUCLEIC ACID EXTRACTION Procedure and/or instrumentation	E	
Name of kit and details of any modifications	E	
Source of additional reagents used Details of DNase or RNAse treatment	D <b>E</b>	
Contamination assessment (DNA or RNA)	E	
Nucleic acid quantification Instrument and method	E E	
Purity (A260/A280)	D	
Yield RNA integrity method/instrument	D E	
RIN/RQI or Cq of 3¹ and 5¹ transcripts Electrophoresis traces	<b>E</b> D	
Inhibition testing (Cq dilutions, spike or other)	E E	
REVERSE TRANSCRIPTION Complete reaction conditions	E	
Amount of RNA and reaction volume	E	
Priming oligonucleotide (if using GSP) and concentration  Reverse transcriptase and concentration	E E	
Temperature and time	E	
Manufacturer of reagents and catalogue numbers  Cqs with and without RT	D D*	
Storage conditions of cDNA	D	
qPCR TARGET INFORMATION  If multiplex, efficiency and LOD of each assay.	E	
Sequence accession number Location of amplicon	<b>E</b> D	
Amplicon length	E	
In silico specificity screen (BLAST, etc) Pseudogenes, retropseudogenes or other homologs?	<b>E</b> D	
Sequence alignment	D	
Secondary structure analysis of amplicon Location of each primer by exon or intron (if applicable)	D E	
What splice variants are targeted?	E	
What splice variants are targeted? qPCR OLIGONUCLEOTIDES Primer sequences	E	
What splice variants are targeted?  qPCR OLIGONUCLEOTIDES  Primer sequences  RTPrimerDB Identification Number	<b>E</b> D	
What splice variants are targeted? qPCR OLIGONUCLEOTIDES Primer sequences RTPrimerDB Identification Number Probe sequences Location and identity of any modifications	E D D** E	
What splice variants are targeted?  qPCR OLIGONUCLEOTIDES  Primer sequences  RTPrimerDB Identification Number  Probe sequences  Location and identity of any modifications  Manufacturer of oligonucleotides  Purification method	<b>E</b> D D**	
What splice variants are targeted? qPCR OLIGONUCLEOTIDES  Primer sequences RTPrimerDB Identification Number Probe sequences Location and identity of any modifications Manufacturer of oligonucleotides Pruffication method qPCR PROTOCOL	E D D*** E D D	
What splice variants are targeted? qPCR OLIGONUCLEOTIDES  Primer sequences RTPrimerDB Identification Number Probe sequences Location and identity of any modifications Manufacturer of oligonucleotides Purification method qPCR PROTOCOL Complete reaction conditions Reaction volume and amount of cDNA/DNA	E D D** E D D	
What splice variants are targeted? qPCR OLIGONUCLEOTIDES  Primer sequences  RTPrimerDB Identification Number  Probe sequences  Location and identity of any modifications  Manufacturer of oligonucleotides  Purification method qPCR PROTOCOL  Complete reaction conditions  Reaction volume and amount of cDNA/DNA  Primer, (probe), Mg++ and dNTP concentrations	E D D*** E D D E E E E	
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What splice variants are targeted?  qPCR OLIGONUCLEOTIDES  Primer sequences  RTPrimerDB Identification Number  Probe sequences  Location and identity of any modifications  Manufacturer of oligonucleotides  Purification method  qPCR PROTOCOL  Complete reaction conditions  Reaction volume and amount of cDNA/DNA  Primer, (probe), Mg++ and dNTP concentrations  Polymerase identity and concentration	E D D D D D D D D D D D D D D E E E E E	
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What splice variants are targeted?  qPCR OLIGONUCLEOTIDES  Primer sequences  RTPrimerDB Identification Number  Probe sequences  Location and identity of any modifications  Manufacturer of oligonucleotides  Purification method  qPCR PROTOCOL  Complete reaction conditions  Reaction volume and amount of cDNA/DNA  Primer, (probe), Mg++ and dNTP concentrations  Polymerase identity and concentration  Buffer/kit identity and manufacturer  Exact chemical constitution of the buffer  Additives (SYBR Green I, DMSO, etc.)  Manufacturer of plates/tubes and catalog number  Complete thermocycling parameters  Reaction setup (manual/robotic)	E D D D E E E D D E E D D D E E D D D D	
What splice variants are targeted? qPCR OLIGONUCLEOTIDES  Primer sequences RTPrimer DB Identification Number Probe sequences Location and identity of any modifications Manufacturer of oligonucleotides Purification method qPCR PROTOCOL Complete reaction conditions Reaction volume and amount of cDNA/DNA Primer, (probe), Mg++ and dNTP concentrations Polymerase identity and concentration Buffer/kit identity and manufacturer Exact chemical constitution of the buffer Additives (SYBR Green I, DMSO, etc.) Manufacturer of plates/tubes and catalog number Complete thermocycling parameters Reaction setup (manual/robotic) Manufacturer of qPCR instrument	E D D D E E E D D D E E E E E E E E E E	
What splice variants are targeted? qPCR OLIGONUCLEOTIDES  Primer sequences RTPrimerDB Identification Number Probe sequences Location and identity of any modifications Manufacturer of oligonucleotides Purification method qPCR PROTOCOL Complete reaction conditions Reaction volume and amount of cDNA/DNA Primer, (probe), Mg++ and dNTP concentrations Polymerase identity and concentration Bufferklt identity and manufacturer Exact chemical constitution of the buffer Additives (SYBR Green I, DMSO, etc.) Manufacturer of plates/tubes and catalog number Complete thermocycling parameters Reaction setup (manual/robotic) Manufacturer of qPCR instrument qPCR VALIDATION Evidence of optimisation (from gradients)	E D D D E E E D D E E D D E E D D D E E D	
What splice variants are targeted? qPCR OLIGONUCLOTIDES  Primer sequences  RTPrimer Sequences  RTPrimer B Identification Number  Probe sequences  Location and identity of any modifications  Manufacturer of oligonucleotides  Purification method qPCR PROTOCOL  Complete reaction conditions  Reaction volume and amount of cDNA/DNA  Primer, (probe), Mg++ and dNTP concentrations  Polymerase identity and concentration  Buffer/kit identity and manufacturer  Exact chemical constitution of the buffer  Additives (SYBR Green I, DMSO, etc.)  Manufacturer of plates/tubes and catalog number  Complete thermocycling parameters  Reaction setup (manual/robotic)  Manufacturer of qPCR instrument qPCR VALIDATION	E D D D D E E D D E E D D E E	
What splice variants are targeted? qPCR OLIGONUCLEOTIDES  Primer sequences RTPrimerDB Identification Number Probe sequences Location and identity of any modifications Manufacturer of oligonucleotides Purification method qPCR PROTOCOL Complete reaction conditions Reaction volume and amount of cDNA/DNA Primer, (probe), Mg++ and dNTP concentrations Polymerase identity and concentration Polymerase identity and concentration Buffer/kit identity and manufacturer Exact chemical constitution of the buffer Additives (SYBR Green I, DMSO, etc.) Manufacturer of plates/tubes and catalog number Complete thermocycling parameters Reaction setup (manual/robotic) Manufacturer of qPCR instrument qPCR VALIDATION Fivience of optimisation (from gradients) Specificity (gel, sequence, melt, or digest) For SYBR Green I, Cq of the NTC Standard curves with slope and y-intercept	E D D D E E D D E E E E E E E E E E E E	
What splice variants are targeted? qPCR OLIGONUCLOTIDES  Primer sequences  RTPrimerDB Identification Number  Probe sequences  Location and identity of any modifications  Manufacturer of oligonucleotides  Purification method qPCR PROTOCOL  Complete reaction conditions  Reaction volume and amount of cDNA/DNA Primer, Grobe), Mg++ and dNTP concentrations  Polymerase identity and concentration  Buffer/kit identity and manufacturer  Exact chemical constitution of the buffer  Additives (SYBR Green J, DMSO, etc.)  Manufacturer of plates/tubes and catalog number  Complete thermocycling parameters  Reaction setup (manual/robotic)  Manufacturer of qPCR instrument qPCR VALIDATION  Evidence of optimisation (from gradients)  Specificity (gel, sequence, melt, or digest)  For SYBR Green J, Cq of the NTC  Standard curves with slope and y-intercept PCR efficiency calculated from slope  Confidence interval for PCR efficiency or standard error	E D D D E E D D E E E E E E E E D D E E E E E E E D D E E E E E E D D E E E D D E	
What splice variants are targeted? qPCR OLIGONUCLEOTIDES  Primer sequences RTPrimerDB Identification Number Probe sequences Location and identity of any modifications Manufacturer of oligonucleotides Purification method qPCR PROTOCOL Complete reaction conditions Reaction volume and amount of cDNA/DNA Primer, (probe), Mg++ and dNTP concentrations Polymerase identity and concentration Buffer/kit identity and manufacturer Exact chemical constitution of the buffer Additives (SYBR Green I, DMSO, etc.) Manufacturer of plates/tubes and catalog number Complete thermocycling parameters Reaction setup (manual/robotic) Manufacturer of qPCR instrument qPCR VALIDATION Evidence of optimisation (from gradients) Specificity (gel, sequence, melt, or digest) For SYBR Green I, Cq of the NTC Standard curves with slope and y-intercept PCR efficiency calculated from slope Confidence interval for PCR efficiency or standard error r2 of standard curve	E D D E E E E E E E E E E D D E E E E E	
What splice variants are targeted? qPCR OLIGONUCLEOTIDES  Primer sequences RTPrimerDB Identification Number Probe sequences Location and identity of any modifications Manufacturer of oligonucleotides Purification method qPCR PROTOCOL Complete reaction conditions Reaction volume and amount of cDNA/DNA Primer, (probe), Mg++ and dNTP concentrations Polymerase identity and concentration Bufferklt identity and manufacturer Exact chemical constitution of the buffer Additives (SyBR Green I, DMSO, etc.) Manufacturer of plates/tubes and catalog number Complete thermocycling parameters Reaction setup (manual/robotic) Manufacturer of qPCR instrument qPCR VALIDATION Evidence of optimisation (from gradients) Specificity (gel, sequence, melt, or digest) For SYBR Green I, Ca of the NTC Standard curves with slope and y-intercept PCR efficiency calculated from slope Confidence interval for PCR efficiency or standard error 12 of standard curve Linear dynamic range Cq variation at lower limit	E D D E E E E E E E E E E E E E E E E E	
What splice variants are targeted? qPCR OLIGONUCLEOTIDES Primer Sequences RTPrimerDB Identification Number Probe sequences Location and identity of any modifications Manufacturer of oligonucleotides Purification method qPCR PROTOCOL Complete reaction conditions Reaction volume and amount of cDNA/DNA Primer, (probe), Mg++ and dNTP concentrations Polymerase identity and concentration Buffer/kit identity and manufacturer Exact chemical constitution of the buffer Additives (SYBR Green I, DMSO, etc.) Manufacturer of plates/tubes and catalog number Complete thermocycling parameters Reaction setup (manual/robotic) Manufacturer of qPCR PKI instrument qPCR VAUIDATION Evidence of optimisation (from gradients) Specificity (leg), sequence, melt, or digest) For SYBR Green I, Cq of the NTC Standard curves with slope and y-intercept PCR efficiency calculated from slope Confidence interval for PCR efficiency or standard error r2 of standard curve Linear dynamic range Cq variation at lower limit Confidence intervals throughout range Evidence for limit of detection	E D D D E E D D E E E E E E E E E E E E	
What splice variants are targeted? qRCR OLIGONUCLEOTIDES  Primer Sequences RTPrimerDB Identification Number Probe sequences Location and identity of any modifications Manufacturer of oligonucleotides Purification method qPCR PROTOCOL Complete reaction conditions Reaction volume and amount of cDNA/DNA Primer, (probe), Mg++ and dNTP concentrations Polymerase identity and concentration Bufferkit identity and manufacturer Exact chemical constitution of the buffer Additives (SyBR Green I, DMSO, etc.) Manufacturer of plates/tubes and catalog number Complete thermocycling parameters Reaction setup (manual/robotic) Manufacturer of qPCR instrument qPCR VALIDATION  Evidence of optimisation (from gradients) Specificity (gel, sequence, melt, or digest) For SYBR Green I, Cq of the NTC Standard curves with slope and y-intercept PCR efficiency calculated from slope Confidence interval for PCR efficiency or standard error r2 of standard curve Linear dynamic range Cq variation at lower limit Confidence intervals throughout range Evidence for limit of detection If multiplex, efficiency and LOD of each assay.	E D D E E E E E E E E E E E E E E E E E	
qPCR OLIGONUCLEOTIDES Primer sequences RTPrimerDB Identification Number Probe sequences Location and identity of any modifications Manufacturer of oligonucleotides Purification method qPCR PROTOCOL Complete reaction conditions Reaction volume and amount of cDNA/DNA Primer, (probe), Mg++ and dNTP concentrations Polymerase identity and concentration Buffer/kit identity and manufacturer Exact chemical constitution of the buffer Additives (SYBR Green I, DMSO, etc.) Manufacturer of plates/bubes and catalog number Complete thermocycling parameters Reaction setup (manual/robotic) Manufacturer of qPCR instrument qPCR VALIDATION Evidence of optimistation (from gradients) Specificity (gel, sequence, melt, or digest) For SYBR Green I, Cq of the NTC Standard curves with slope and y-intercept PCR efficiency calculated from slope Confidence interval for PCR efficiency or standard error r2 of standard curve Linear dynamic range Cq variation at lower limit Confidence intervals throughout range Evidence for limit of detection If multiplex, efficiency and LOD of each assay, DATA ANALYSIS qPCR analysis program (source, version)	E D D D E E E E E E E E E E E E E E E E	
What splice variants are targeted?  Primer sequences  RTPrimerDB Identification Number  Probe sequences  Location and identity of any modifications  Manufacturer of oligonucleotides  Purification method  Purification method  Purification noditions  Reaction volume and amount of CDNA/DNA  Primer, (probe), Mg++ and dNTP concentrations  Polymerase identity and concentration  Bufferkit identity and manufacturer  Exact chemical constitution of the buffer  Additives (SYBR Green I, DMSO, etc.)  Manufacturer of plates/tubes and catalog number  Complete thermocycling parameters  Reaction setup (manual/robotic)  Manufacturer of optimisation (from gradients)  Specificity (gel, sequence, melt, or digest)  For SYBR Green I, Cq of the NTC  Standard curves with slope and y-intercept  PCR efficiency calculated from slope  Confidence interval for PCR efficiency or standard error  r2 of standard curve  Linear dynamic range  Cq variation at lower limit  Confidence intervals throughout range  Evidence for limit of detection  If multiplex, efficiency and LOD of each assay.  DATA NANAYSIS  QPCR analysis program (source, version)  Cq method determination	E D D E E E E E E E E E E E E E E E E E	
What splice variants are targeted?  Primer Sequences  RTPrimerDB Identification Number  Probe sequences  Location and identity of any modifications  Manufacturer of oligonucleotides  Purification method  Purification method  QPCR PROTOCOL  Complete reaction conditions  Reaction volume and amount of cDNA/DNA  Primer, (probe), Mg++ and dNTP concentrations  Polymerase identity and concentration  Bulferkit identity and concentration  Bulferkit identity and manufacturer  Exact chemical constitution of the buffer  Additives (SYBR Green I, DMSO, etc.)  Manufacturer of plates/tubes and catalog number  Complete thermocycling parameters  Reaction setup (manual/robotic)  Manufacturer of qPCR instrument  QPCR VALIDATION  Evidence of optimisation (from gradients)  Specificity (gel, sequence, melt, or digest)  For SYBR Green I, Cq of the NTC  Standard curves with slope and y-intercept  PCR efficiency calculated from slope  Confidence interval for PCR efficiency or standard error  r2 of standard curve  Linear dynamic range  Cq variation at lower limit  Confidence intervals throughout range  Evidence for limit of detection  If multiplex, efficiency and LOD of each assay.  DAYA ANALYSIS  QPCR analysis program (source, version)  Cq method determination  Outlier identification and disposition	E D D E E E E E E E E E E E E E E E E E	
What splice variants are targeted?  Primer sequences  RTPrimerDB Identification Number Probe sequences Location and identity of any modifications Manufacturer of oligonucleotides Purification method  QPCR PROTOCOL  Complete reaction conditions Reaction volume and amount of cDNA/DNA Primer, (probe), Mg++ and dNTP concentrations Polymerase identity and concentration Buffer/kit identity and manufacturer Exact chemical constitution of the buffer Additives (SYBR Green I, DMSO, etc.) Manufacturer of plates/tubes and catalog number Complete themocycling parameters Reaction setup (manual/robotic) Manufacturer of qPCR instrument QPCR VALIDATION  Evidence of optimisation (from gradients) Specificity (gel, sequence, melt, or digest) For SYBR Green I, Cq of the NTC Slandard curves with slope and y-intercept PCR Grificiency calculated from slope Confidence intervals for PCR efficiency or standard error r2 of standard curve Linear dynamic range Cq variation at lower limit Confidence intervals throughout range Fividence for limit of detection If multiplex, efficiency and LOD of each assay.  DATA ANALYSIS QPCR analysis program (source, version) Cq method determination Outlier identification and disposition Results of NTCS Instrument and choice of reference genes	E D D E E E E E E E E E E E E E E E E E	
What splice variants are targeted?  gPCR OLIGONUCLEOTIDES  Primer sequences  RTPrimerDB Identification Number  Probe sequences  Location and identity of any modifications  Manufacturer of oligonucleotides  Purification method  gPCR PROTOCOL  Complete reaction conditions  Reaction volume and amount of cDNA/DNA  Primer, (probe), Mg++ and dNTP concentrations  Polymerase identity and concentration  Buffer/kit identity and manufacturer  Exact chemical constitution of the buffer  Additives (SYBR Green I, DMSO, etc.)  Manufacturer of plates/tubes and catalog number  Complete thermocycling parameters  Reaction setur (manual/robotic)  Manufacturer of gPCR instrument  gPCR VAILDATION  Specificity (gel, sequence, melt, or digest)  For SYBR Green I, Cg of the NTC  Standard curves with slope and y-intercept  PCR efficiency calculated from slope  Confidence intervals for PCR efficiency or standard error  r2 of standard curve  Linear dynamic range  Cq variation at lower limit  Confidence intervals throughout range  Evidence for limit of detection  If multiplex, efficiency and LOD of each assay.  DATA ANALYSIS  gPCR analysis program (source, version)  Cq method determination  Outlier identification and disposition  Results of NTCS  Justification of normalisation method  Number and concordance of biological replicates	E D D E E E E E E E E E E E E E E E E E	
What splice variants are targeted?  PCR OLIGONUCLEOTIDES  Primer sequences  RTPrimerDB Identification Number  Probe sequences  Location and identity of any modifications  Manufacturer of oligonucleotides  Purification method  QRC RYROTOCOL  Complete reaction conditions  Reaction volume and amount of cDNA/DNA  Primer, propobe, Mg++ and dNTP concentrations  Polymerase identity and concentration  Buffer/kit identity and manufacturer  Exact chemical constitution of the buffer  Additives (SYBR Green I, DMSO, etc.)  Manufacturer of plates/tubes and catalog number  Complete thermocycling parameters  Reaction setup (manual/robotic)  Manufacturer of qPCR instrument  QRC RVALIDATION  Evidence of optimisation (from gradients)  Specificity (gel, sequence, melt, or digest)  For SYBR Green I, Cq of the NTC  Standard curves with slope and y-intercept  PCR efficiency calculated from slope  Confidence interval for PCR efficiency or standard error  r2 of standard curve  Linear dynamic range  Cq variation at lower limit  Confidence intervals throughout range  Evidence for limit of detection  If multiplex, efficiency and LOD of each assay,  DYANAMISIS  QPCR analysis program (source, version)  Cq method determination  Outlier identification and disposition  Results of NTCS  Lustification of number and choice of reference genes  Description of normalisation method  Number and concordance of biological replicates  Number and stage (RT or QPCR) of technical replicates	E D D D E E E E E E E E E E E E E E E E	
What splice variants are targeted?  PCR OLICONUCLEOTIDES  Primer sequences  RIPrimerDB Identification Number  Probe sequences  Location and identity of any modifications  Manufacturer of oligonucleotides  Purification method  PCR PROTOCOL  Complete reaction conditions  Reaction volume and amount of cDNA/DNA  Primer, Iprobo, Mgx++ and dNTP concentrations  Polymerase identity and concentration  BufferRit identity and manufacturer  Exact chemical constitution of the buffer  Additives (SYBR Green I, DMSO, etc.)  Manufacturer of plates/tubes and catalog number  Complete thermocycling parameters  Reaction setup (manual/robotic)  Manufacturer of plates/tubes and catalog number  Complete of primination of the buffer  Additives (SyBR Green I, DMSO, etc.)  Manufacturer of plates/tubes and catalog number  Complete of optimisation (from gradients)  Specificity (gel, sequence, melt, or digest)  For SYBR Green I, Cq of the NTC  Standard curves with slope and v-intercept  PCR efficiency calculated from slope  Confidence interval for PCR efficiency or standard error  r2 of standard curve  Linear dynamic range  Cq variation at lower limit  Confidence intervals throughout range  Evidence for limit of detection  If multiplex, efficiency and LOD of each assay.  DATA ANAINSIS  DATA ANAINSIS  DATA ANAINSIS  Unuber and choice of reference genes  Description of normalisation method  Number and concordance of biological replicates  Repoalability (inter-assay variation), %CV)	E D D E E E E E E E E E E E E E E E E E	
What splice variants are targeted?  gPCR OLICONUCLEUTIDES  Primer sequences  RTPrimerDB Identification Number  Probe sequences  Location and identity of any modifications  Manufacturer of oligonucleotides  Purification method  gPCR PROTOCOL  Complete reaction conditions  Reaction volume and amount of cDNA/DNA  Primer, (proble), Mg++ and dNTP concentrations  Polymerase identity and concentration  Buffer/Rt identity and manufacturer  Exact chemical constitution of the buffer  Additives (SYBR Green I, DMSO, etc.)  Manufacturer of plates/tubes and catalog number  Complete thermocycling parameters  Reaction setup (manual/robotic)  Manufacturer of qPCR instrument  gPCR VALIDATION  Evidence of optimisation (from gradients)  Specificity (gel, sequence, melt, or digest)  For SYBR Green I, Cg of the NTC  Standard curves with slope and y-intercept  PCR efficiency calculated from slope  Confidence interval for PCR efficiency or standard error  12 of standard curve  Linear dynamic range  Cq variation at lower limit  Confidence interval strong mounts of the saw.  DATA Analysis  gPCR analysis program (source, version)  Cq method determination  Outlier identification and disposition  Results of NTCS  Iustification of number and choice of reference genes  Description of normalisation method  Number and stage (RT or gPCR) of technical replicates  Number and stage (RT or gPCR) of technical replicates  Number and stage (RT or gPCR) of technical replicates  Number and stage (RT or gPCR) of technical replicates  Number and stage (RT or gPCR) of technical replicates  Number and stage (RT or gPCR) of technical replicates  Number and stage (RT or gPCR) of technical replicates	E D D E E E E E E E E E E E E E E E E E	
What splice variants are targeted?  PCR OLICONUCLEOTIDES  Primer sequences  RTPrimerDB Identification Number  Probe sequences  Location and identity of any modifications  Manufacturer of oligonucleotides  Purification method  QPCR PROTOCOL  Complete reaction conditions  Reaction volume and amount of cDNA/DNA  Primer, (probe), Mg++ and dNTP concentrations  Polymerase identity and concentration  Bufferkit identity and manufacturer  Exact chemical constitution of the buffer  Additives (SYBR Green I, DMSO, etc.)  Manufacturer of plates/tubes and catalog number  Complete thermocycling parameters  Reaction setup (manual/robotic)  Manufacturer of qPCR instrument  QPCR VALIDATION  Evidence of optimisation (from gradients)  Specificity (gel, sequence, melt, or digest)  For SYBR Green I, Cq of the NTC  Standard curves with slope and y-intercept  PCR efficiency calculated from slope  Confidence intervals throughout range  Evidence for limit of detection  If multiplex, efficiency and LOD of each assay.  DATA NALYSIS  QPCR analysis program (source, version)  Cq method determination  Outlier identification and disposition  Results of NTICs  Iustification of number and choice of reference genes  Description of normalisation method  Number and sonocraches of biological replicates  Number and stage (RT or qPCR) of technical replicates  Number and stages (RT or qPCR) of technical replicates  Number and stages (RT or qPCR) of technical replicates  Number and stages (RT or qPCR) of technical replicates  Number and stage (RT or qPCR) of technical replicates  Number and stages (RT or qPCR) of technical replicates  Number and stages (RT or qPCR) of technical replicates  Number and stages (RT or qPCR) of technical replicates  Number and stages (RT or qPCR) of technical replicates  Number and stages (RT or qPCR) of technical replicates  Number and stages (RT or qPCR) of technical replicates  Number and stages (RT or qPCR) of technical replicates	E D D E E E E E E E E E E E E E E E E E	

**Table 1.** MIQE checklist for authors, reviewers and editors. All essential information (E) must be submitted with the manuscript. Desirable information (D) should be submitted if available. If using primers obtained from RTPrimerDB, information on qPCR target, oligonucleotides, protocols and validation is available from that source.

<sup>\*:</sup> Assessing the absence of DNA using a no RT assay is essential when first extracting RNA. Once the sample has been validated as RDNA-free, inclusion of a no-RT control is desirable, but no longer essential.

<sup>\*\*:</sup> Disclosure of the probe sequence is highly desirable and strongly encouraged. However, since not all commercial pre-designed assay vendors provide this information, it cannot be an essential requirement. Use of such assays is advised against.