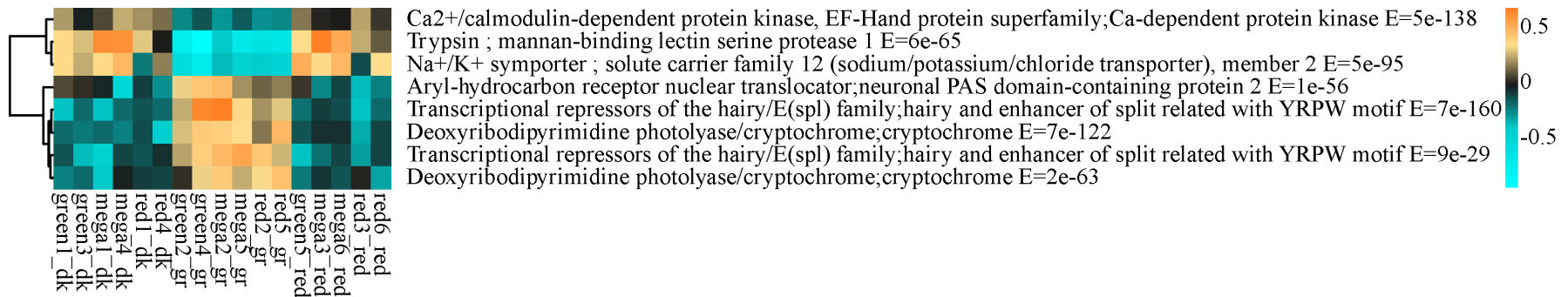
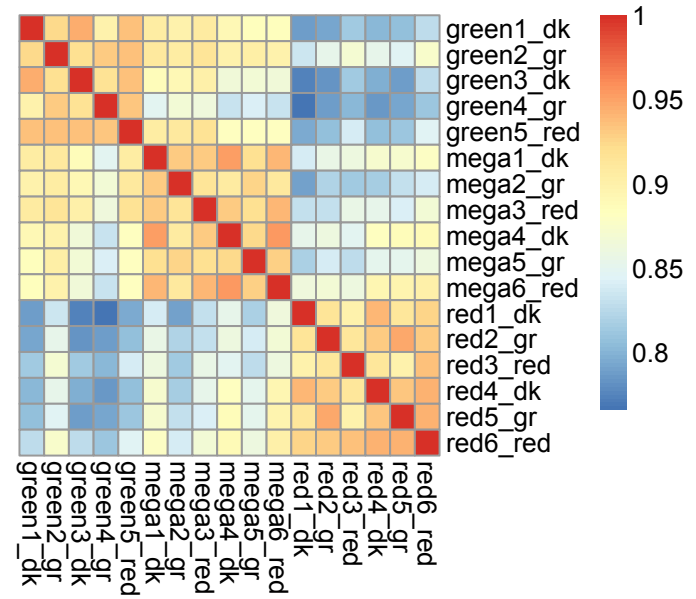


Nice R package for making heatmaps

pheatmap



[http://cran.r-project.org/
web/packages/pheatmap/
index.html](http://cran.r-project.org/web/packages/pheatmap/index.html)



```

> head(exp)
      green1_dk mega1_dk green2_gr mega2_gr mega3_red green3_dk
isogroup10979 6.296149 6.570837 5.899552 5.356431 6.724022 6.417827
isogroup13244 6.750216 7.029649 5.562750 5.938274 7.087965 6.659479
isogroup1996  7.139482 7.142619 6.171526 6.373726 7.148896 7.033077
isogroup7904  5.356431 5.356431 6.035024 6.498003 5.685713 5.577790
isogroup3676  6.211784 5.871480 6.621497 6.867575 6.154541 5.978316
isogroup29627 5.356431 5.871480 5.356431 5.738803 5.356431 5.356431

      mega4_dk green4_gr mega5_gr green5_red mega6_red red1_dk
isogroup10979 6.772880 5.916744 5.803765 6.391164 6.483878 6.556468
isogroup13244 7.013566 5.356431 5.615388 6.728181 6.925941 6.689225
isogroup1996  7.263313 5.975031 6.275734 7.300427 7.267887 6.527665
isogroup7904  5.578430 6.457216 6.165769 5.559693 5.645039 5.632151
isogroup3676  6.257922 6.788670 6.936609 5.962260 6.255969 6.216852
isogroup29627 5.356431 5.356431 5.356431 5.356431 5.356431 6.689225

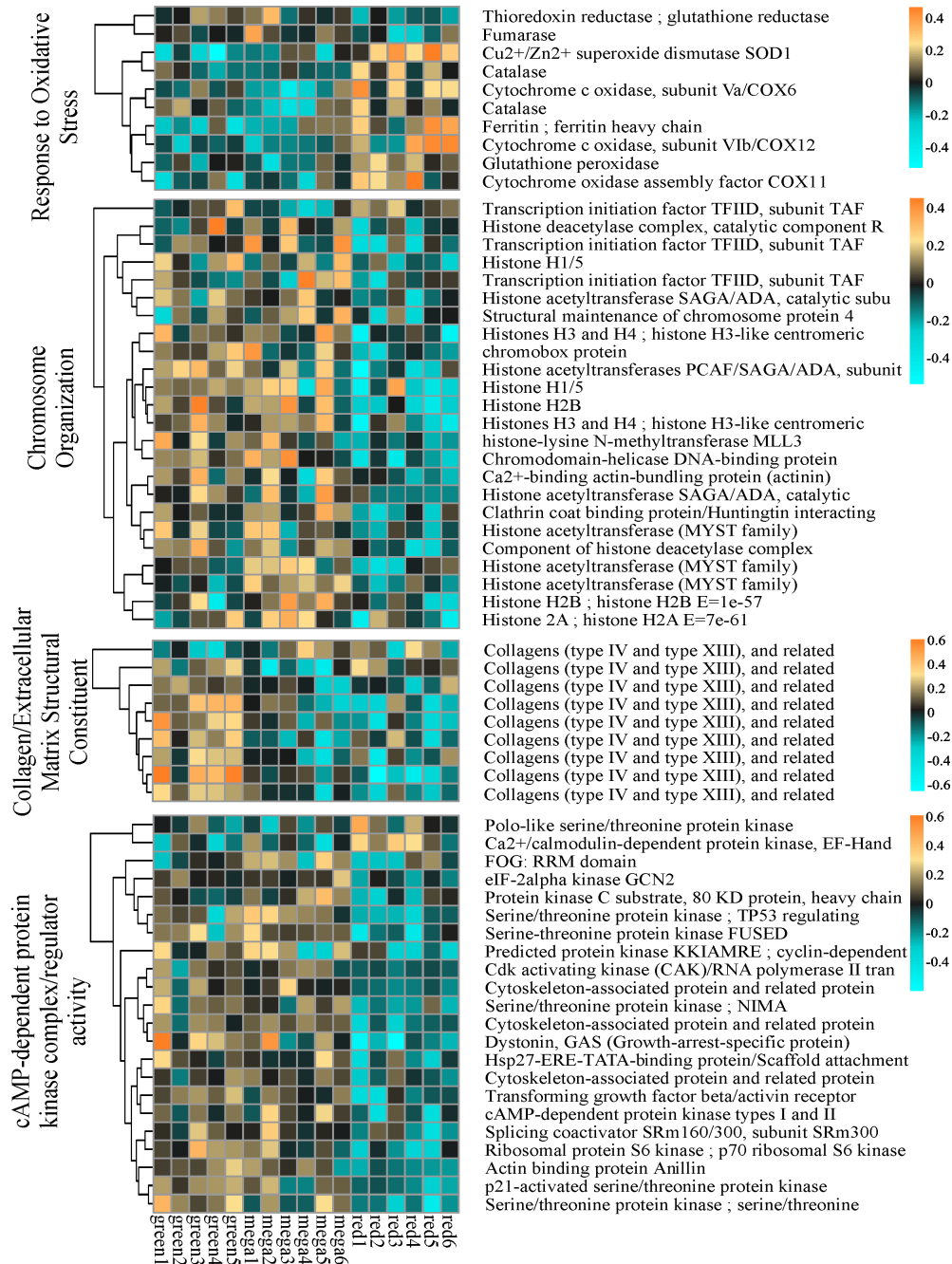
      red2_gr red3_red red4_dk red5_gr red6_red
isogroup10979 5.356431 6.378329 6.654834 5.903847 6.371307
isogroup13244 5.677243 6.638230 6.357824 5.650287 6.485595
isogroup1996  6.328771 6.638230 6.935248 6.249401 7.133316
isogroup7904  5.994173 5.356431 5.574034 6.098703 5.567701
isogroup3676  6.818004 6.045460 6.312892 6.660526 6.263160
isogroup29627 6.376728 6.086411 6.461024 6.335759 6.110251
>

```

>library(pheatmap)

>pheatmap(cor(exp),cluster_rows=F,cluster_cols=F)

>pheatmap(cor(exp))



>means=apply(exp,1,mean)

means of rows

>explc=exp-means

subtracting them

>pdf

("file_name",height=35,width=9)

>pheatmap(as.matrix(explc),color

= colorRampPalette(rev(c

("#D73027", "#D73027", "#D73027

", "#FEE090", "#FFFFBF",

"#91BFDB", "#4575B4", "#4575B4")

))

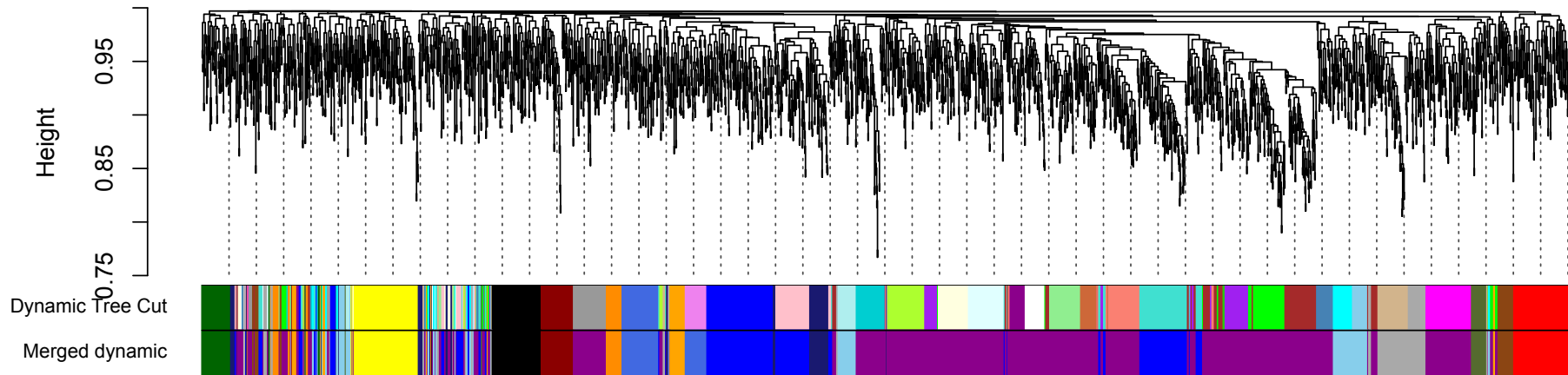
(100),cex=0.9,border_color=NA,cl

ustering_distance_rows="correlati

on",clustering_distance_cols="cor

relation")

>dev.off()



WGCNA

Dept. of Human Genetics, UC Los Angeles (PL, SH), and Dept. of Biostatistics, UC Los Angeles (SH)

Peter (dot) Langfelder (at) gmail (dot) com, SHorvath (at) mednet (dot) ucla (dot) edu

BMC Bioinformatics, 2008 9:559

Special thanks to Rachel Wright for slides

Extensive Tutorials Online

- <http://labs.genetics.ucla.edu/horvath/CoexpressionNetwork/Rpackages/WGCNA/Tutorials/>
- <http://labs.genetics.ucla.edu/horvath/CoexpressionNetwork/Rpackages/WGCNA/>
- R package

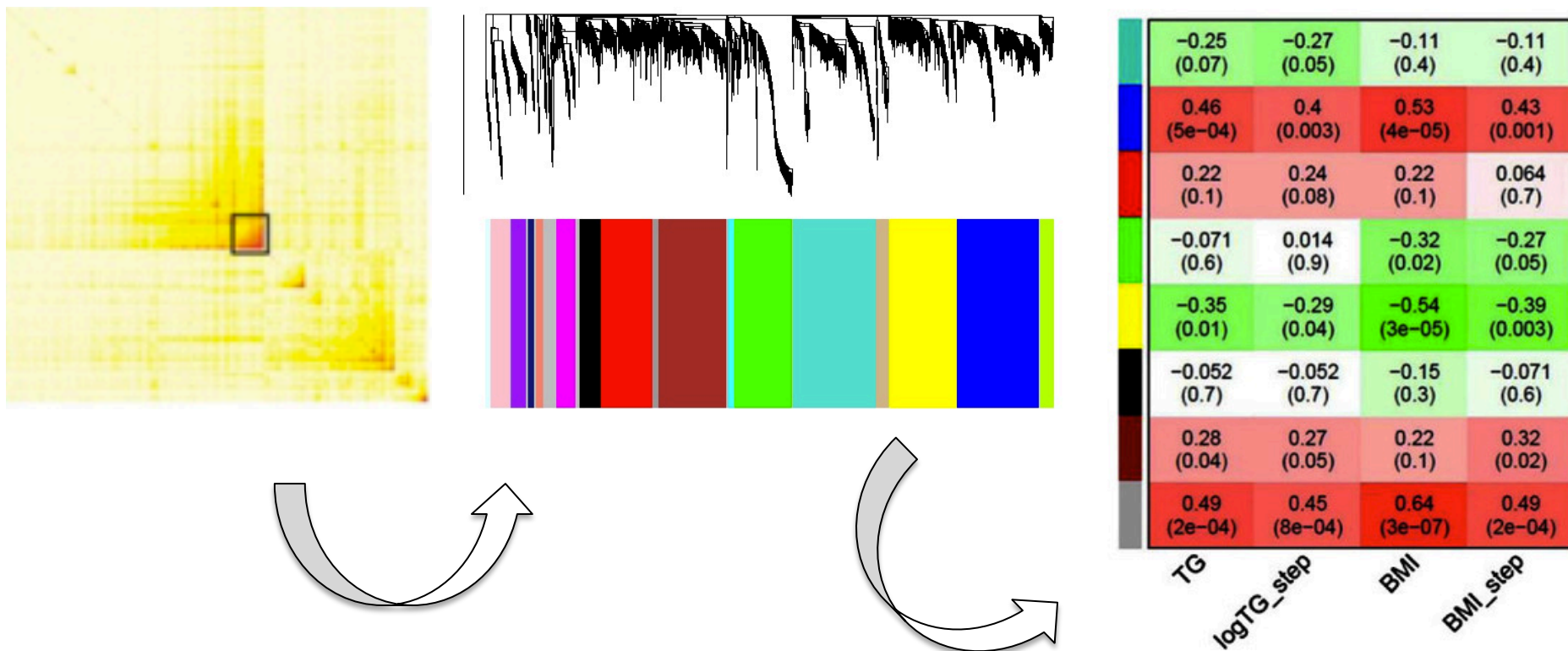
```
>source("http://bioconductor.org/biocLite.R")
```

```
>biocLite("impute")
```

```
>install.packages("WGCNA")
```

WGCNA: weighted gene coexpression network analysis

- Identify groups (modules) of co-regulated genes
- Correlate module expression values to trait data



Benefits of WGCNA

- Unsupervised and unbiased
 - Bypasses multiple testing correction
- Finds unique co-expression modules that correspond to clusters of correlated transcripts

What kind of data do you need?

- Normalized counts data files (.csv)
 - VSD file: variance transformed counts data
 - Generated from DESeq
 - The tutorial you use depends on how large your dataset is (18000 genes? Or 2000 genes?)
- Trait Data
 - Made in excel, saved as .csv

VSD file

- > cdsBlind = estimateDispersions(cds, method="blind")
- > vsd = varianceStabilizingTransformation (cdsBlind)

```
> head(exp)
      green1_dk mega1_dk green2_gr mega2_gr mega3_red green3_dk
isogroup10979 6.296149 6.570837 5.899552 5.356431 6.724022 6.417827
isogroup13244 6.750216 7.029649 5.562750 5.938274 7.087965 6.659479
isogroup1996 7.139482 7.142619 6.171526 6.373726 7.148896 7.033077
isogroup7904 5.356431 5.356431 6.035024 6.498003 5.685713 5.577790
isogroup3676 6.211784 5.871480 6.621497 6.867575 6.154541 5.978316
isogroup29627 5.356431 5.871480 5.356431 5.738803 5.356431 5.356431
      mega4_dk green4_gr mega5_gr green5_red mega6_red red1_dk
isogroup10979 6.772880 5.916744 5.803765 6.391164 6.483878 6.556468
isogroup13244 7.013566 5.356431 5.615388 6.728181 6.925941 6.689225
isogroup1996 7.263313 5.975031 6.275734 7.300427 7.267887 6.527665
isogroup7904 5.578430 6.457216 6.165769 5.559693 5.645039 5.632151
isogroup3676 6.257922 6.788670 6.936609 5.962260 6.255969 6.216852
isogroup29627 5.356431 5.356431 5.356431 5.356431 5.356431 6.689225
      red2_gr red3_red red4_dk red5_gr red6_red
isogroup10979 5.356431 6.378329 6.654834 5.903847 6.371307
isogroup13244 5.677243 6.638230 6.357824 5.650287 6.485595
isogroup1996 6.328771 6.638230 6.935248 6.249401 7.133316
isogroup7904 5.994173 5.356431 5.574034 6.098703 5.567701
isogroup3676 6.818004 6.045460 6.312892 6.660526 6.263160
isogroup29627 6.376728 6.086411 6.461024 6.335759 6.110251
>
```

Trait Data file (upload in R as .csv)

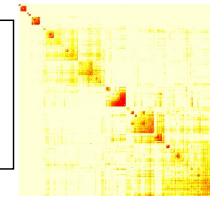
- Categorical or quantitative traits
- Examples:

Sample	mega	green	red	dark_light	green_light	red_light	weight
green1_dk	0	1	0	1	0	0	8
green2_gr	0	1	0	0	1	0	9
green3_dk	0	1	0	1	0	0	10
green4_gr	0	1	0	0	1	0	11.2
green5_red	0	1	0	0	0	1	7
mega1_dk	1	0	0	1	0	0	9
mega2_gr	1	0	0	0	1	0	12.5
mega3_red	1	0	0	0	0	1	8.7
mega4_dk	1	0	0	1	0	0	9.2
mega5_gr	1	0	0	0	1	0	10.2
mega6_red	1	0	0	0	0	1	13
red1_dk	0	0	1	1	0	0	14
red2_gr	0	0	1	0	1	0	6
red3_red	0	0	1	0	0	1	3
red4_dk	0	0	1	1	0	0	2
red5_gr	0	0	1	0	1	0	9.8
red6_red	0	0	1	0	0	1	10

Construct a gene co-expression network

Rationale: make use of interaction patterns among genes

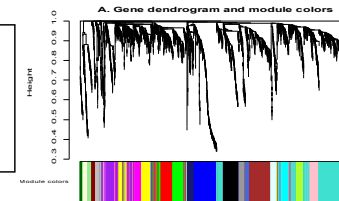
Tools: correlation as a measure of co-expression



Identify modules

Rationale: module (pathway) based analysis

Tools: hierarchical clustering, Dynamic Tree Cut

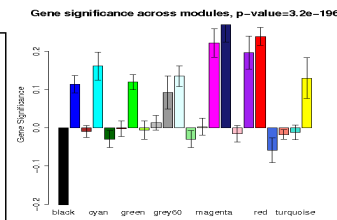


Relate modules to external information

Array Information: clinical data, SNPs, proteomics

Gene Information: ontology, functional enrichment

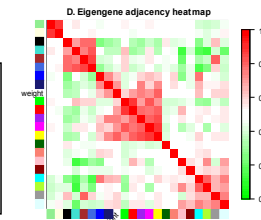
Rationale: find biologically interesting modules



Study module relationships

Rationale: biological data reduction, systems-level view

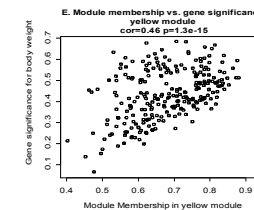
Tools: Eigengene Networks



Find the key drivers in interesting modules

Rationale: experimental validation, biomarkers

Tools: intramodular connectivity, causality testing



Overview of WGCNA methodology. This flowchart presents a brief overview of the main steps of Weighted Gene Co-expression Network Analysis.

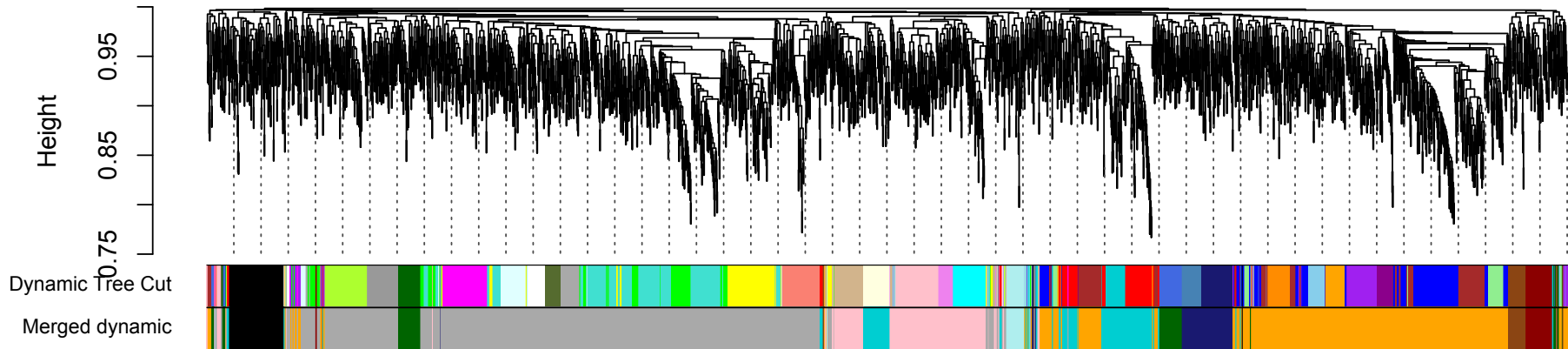
Langfelder and Horvath BMC Bioinformatics 2008 9:559 doi:10.1186/1471-2105-9-559

Step 2: Identifying modules

Dendrogram and gene modules

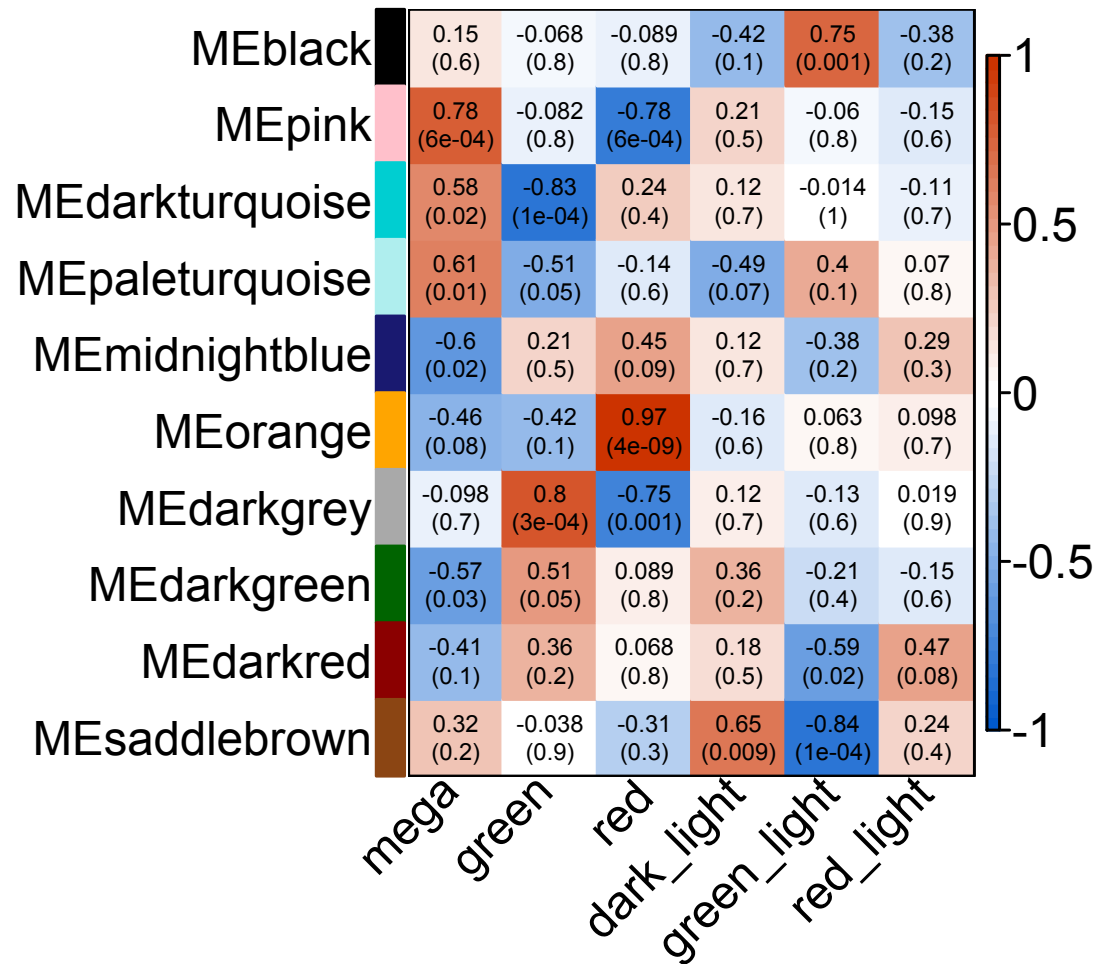
Clustering dendrogram of top host DEGs, with dissimilarities based on topological overlap shown with assigned module colors (Dynamic tree cut) and upon merging modules whose expression profiles were 75% similar (Merged dynamic).

Cluster Dendrogram



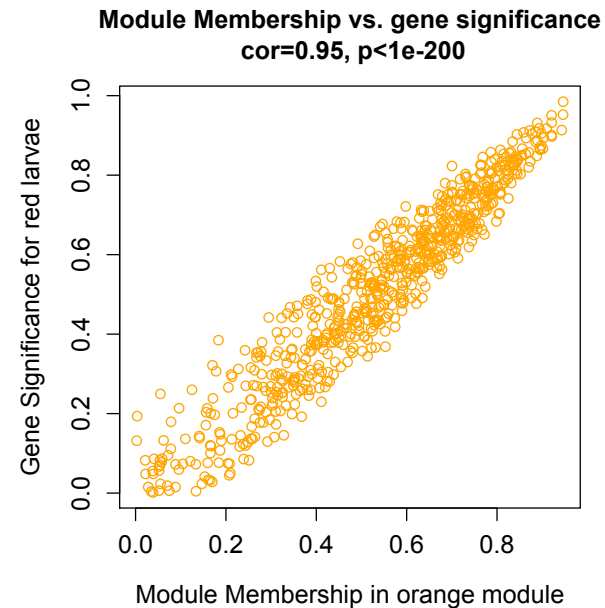
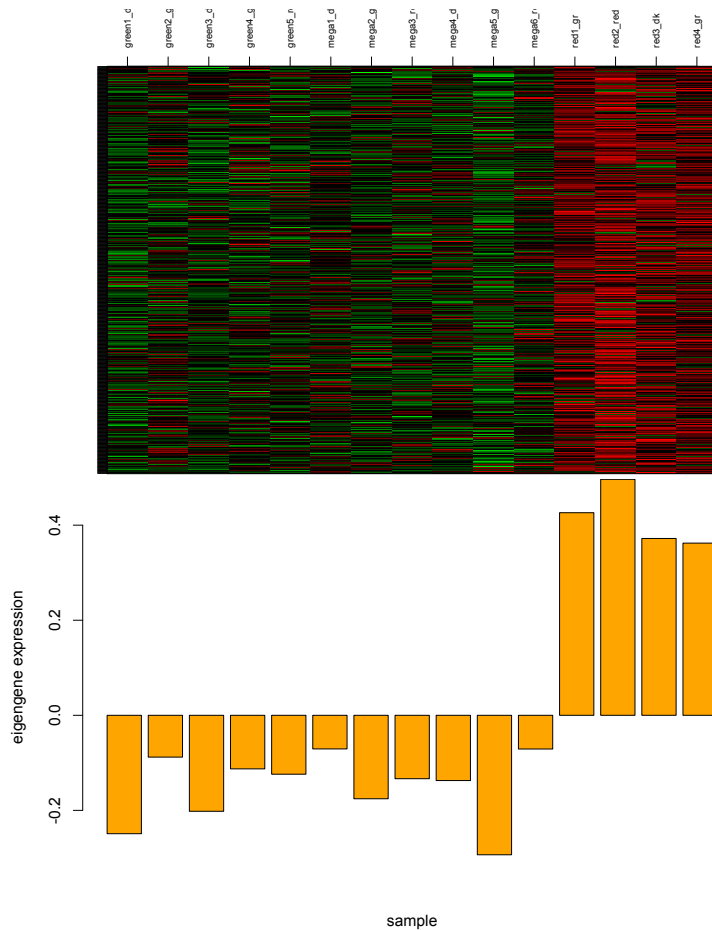
Step 3: Relate Modules to external information

Module-trait relationships



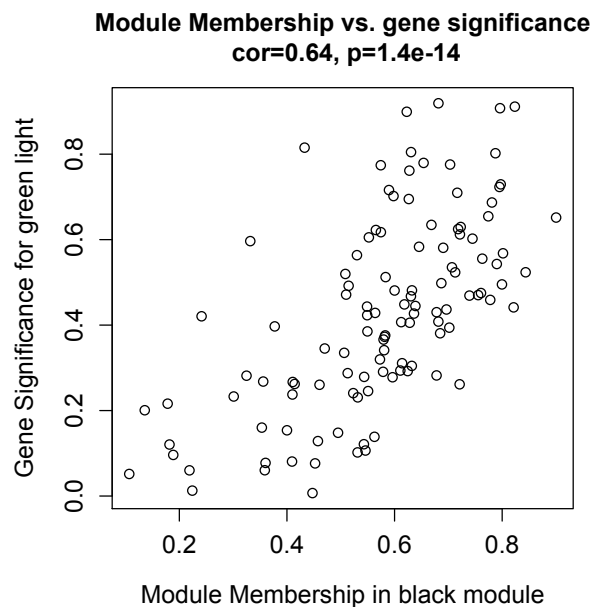
How well do your modules represent your traits?

- Module Eigengene concept
 - Baseline expression in the module
 - “The module eigengene E is defined as the first principal component of a given module. It can be considered a representative of the gene expression profiles in a module.”



kME concept

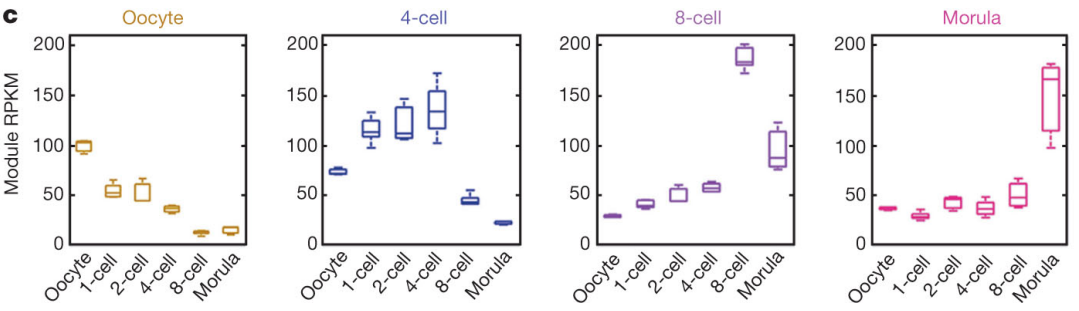
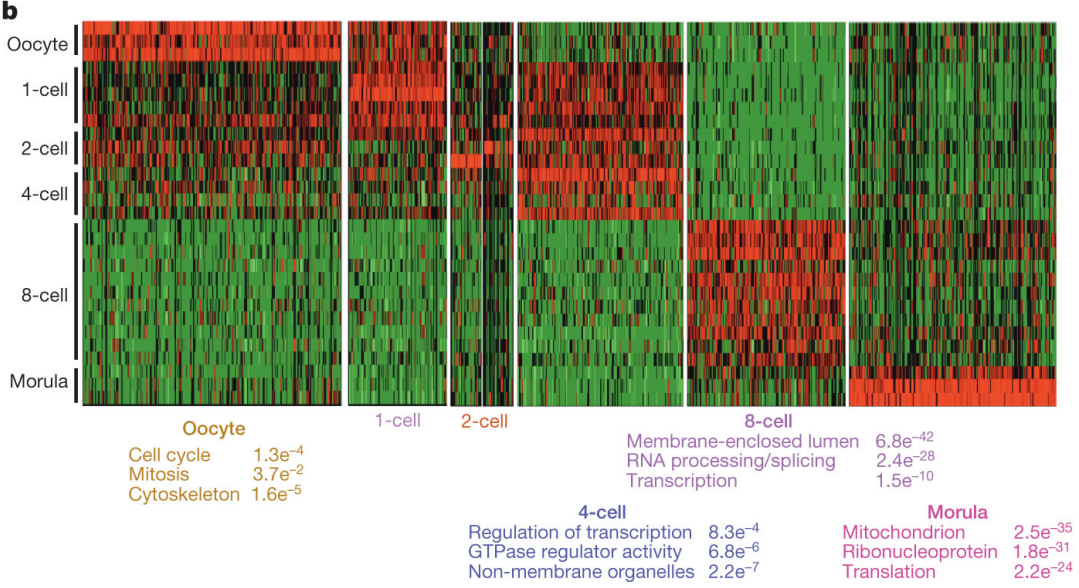
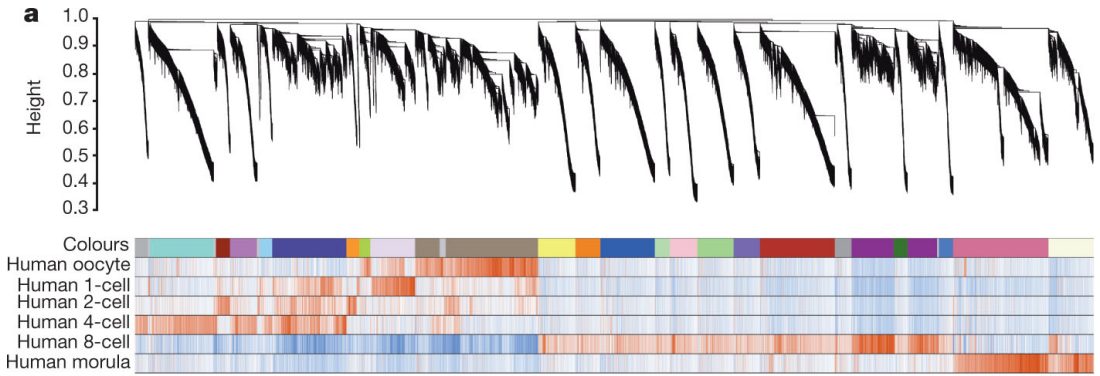
- High kME (MM) values = high correlation to the eigengene
- “Highly connected intramodular hub genes tend to have high module membership values to the respective module.”
- kME values are useful for selecting the “most important” genes within a module
- “Gene significance of 0 indicates that the gene is not significant with regard to the biological question of interest”



An Example:

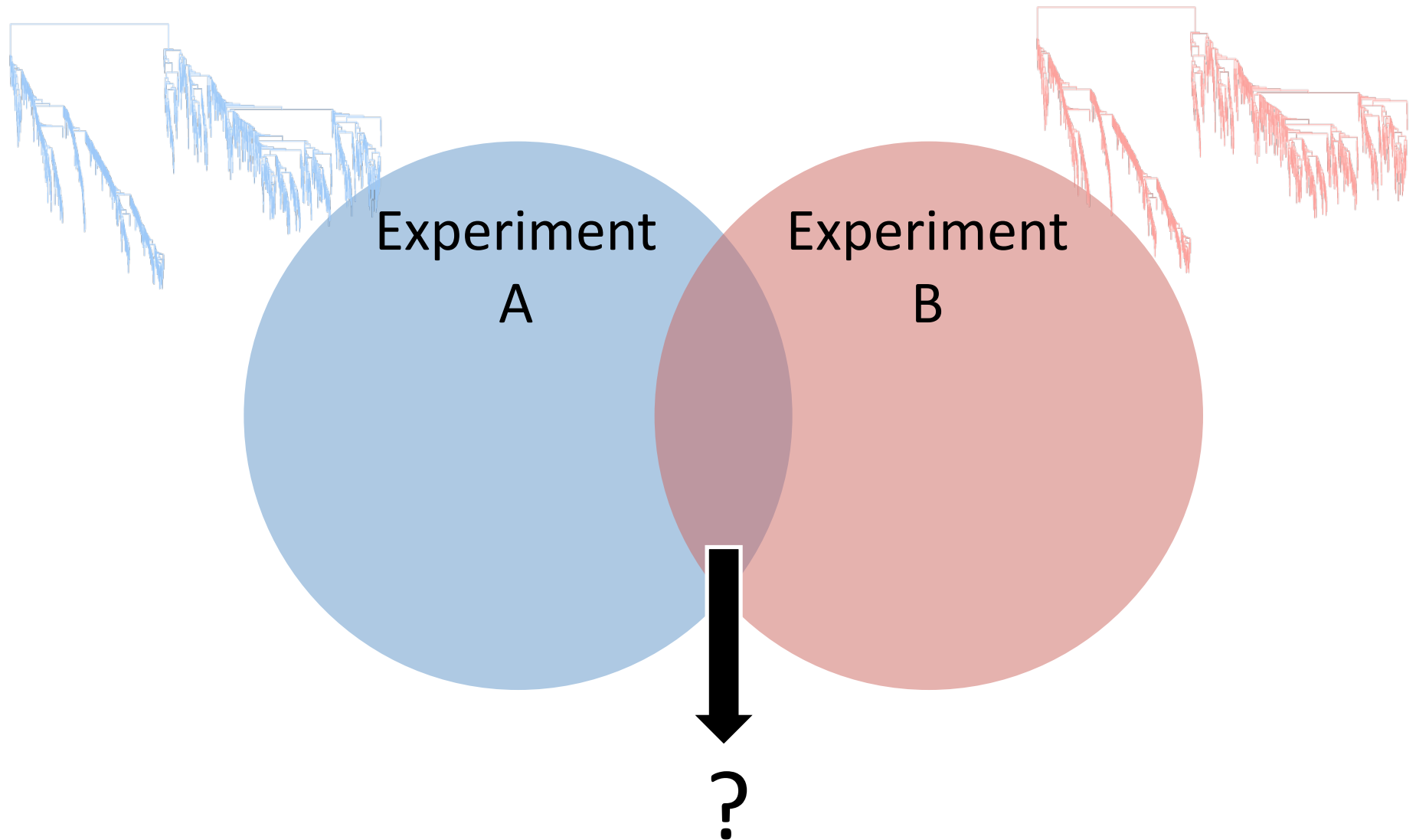
- **Genetic programs in human and mouse early embryos revealed by single-cell RNA sequencing**
 - Zhigang Xue^{1*}, Kevin Huang^{2*} *et al*
- Analysis of transcriptome dynamics from oocyte to morula in both human and mouse embryos
 - Single-cell RNAseq
- They found that each developmental stage is represented by a small number of functional modules of co-expressed genes
- They also identified conserved key members (or ‘hub’ genes) of human and mouse networks that likely drive mammalian pre-implantation development

Network analysis of human pre-implantation development.

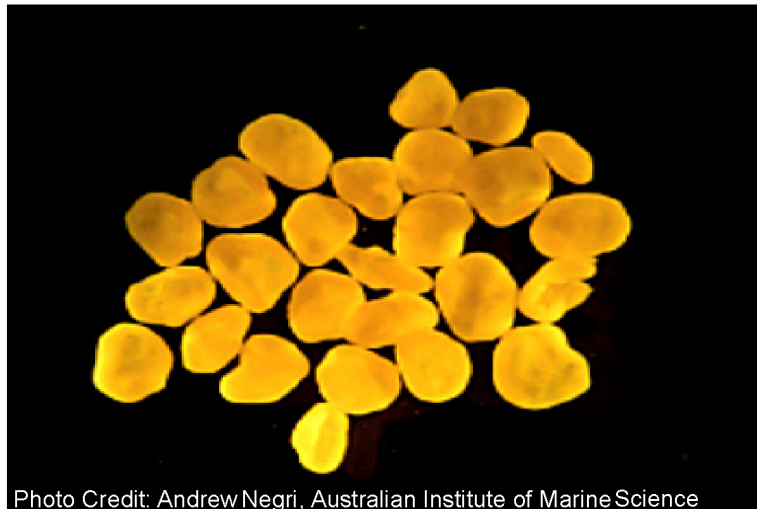


Z Xue *et al.* *Nature* **000**,
1-5 (2013) doi:10.1038/
nature12364

Comparing data sets with module conservation analysis



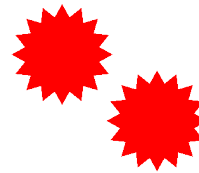
A: Gene expression analysis of *Acropora millepora* larvae challenged with viruses and heat



Mix gametes



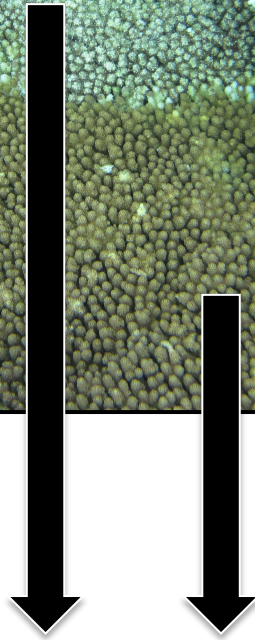
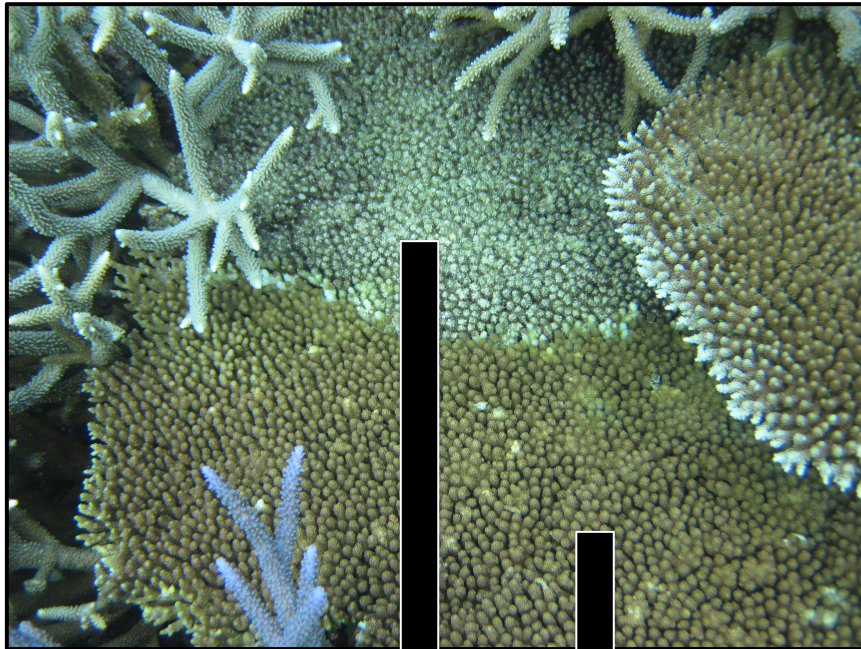
Add VLPs at two-cell stage



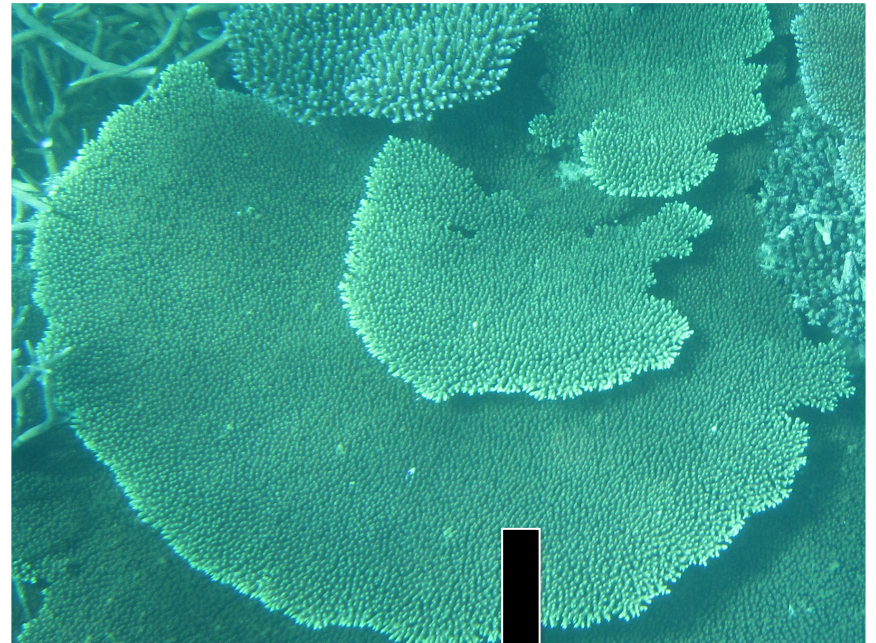
Quantify development through the prawn chip stage

B: Gene expression analysis of adult *Acropora hyacinthus* affected by White Syndrome

Diseased

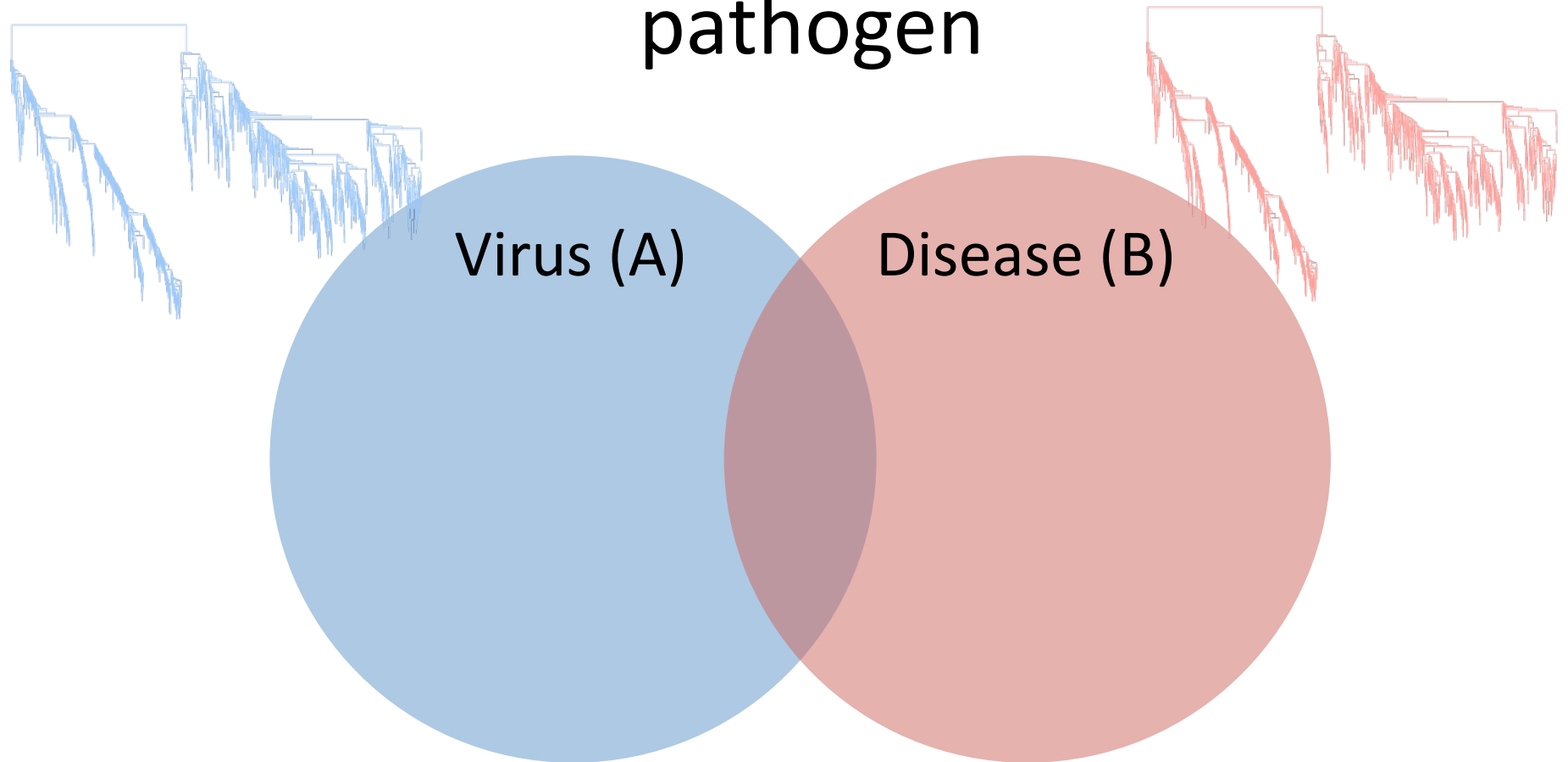


Healthy



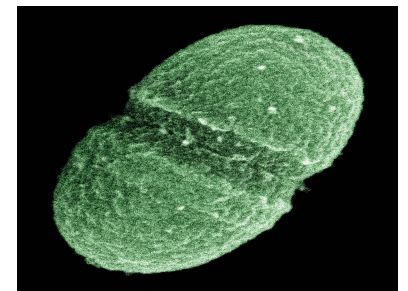
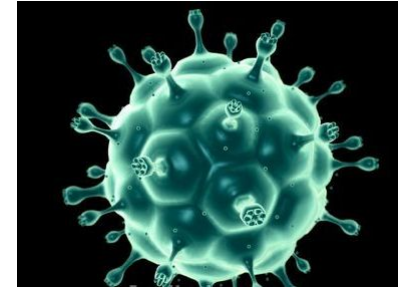
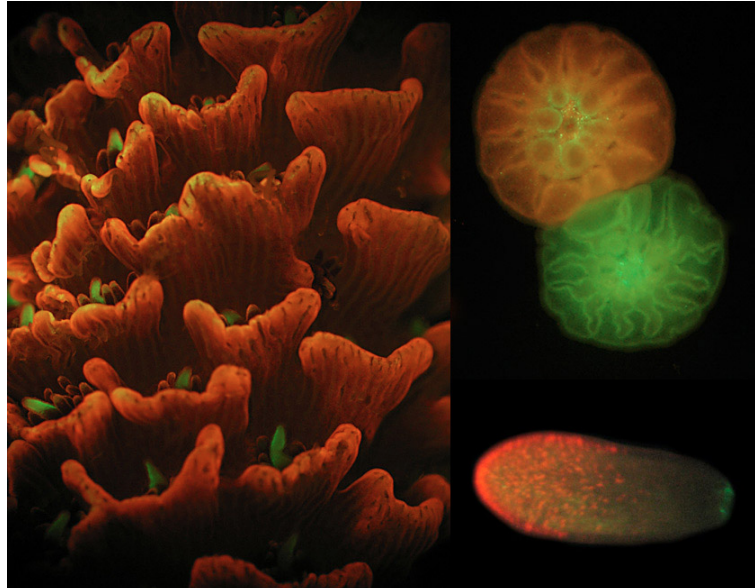
The two experiments are common in
genus and type of stress...

but differ in species, life-stage, and
pathogen

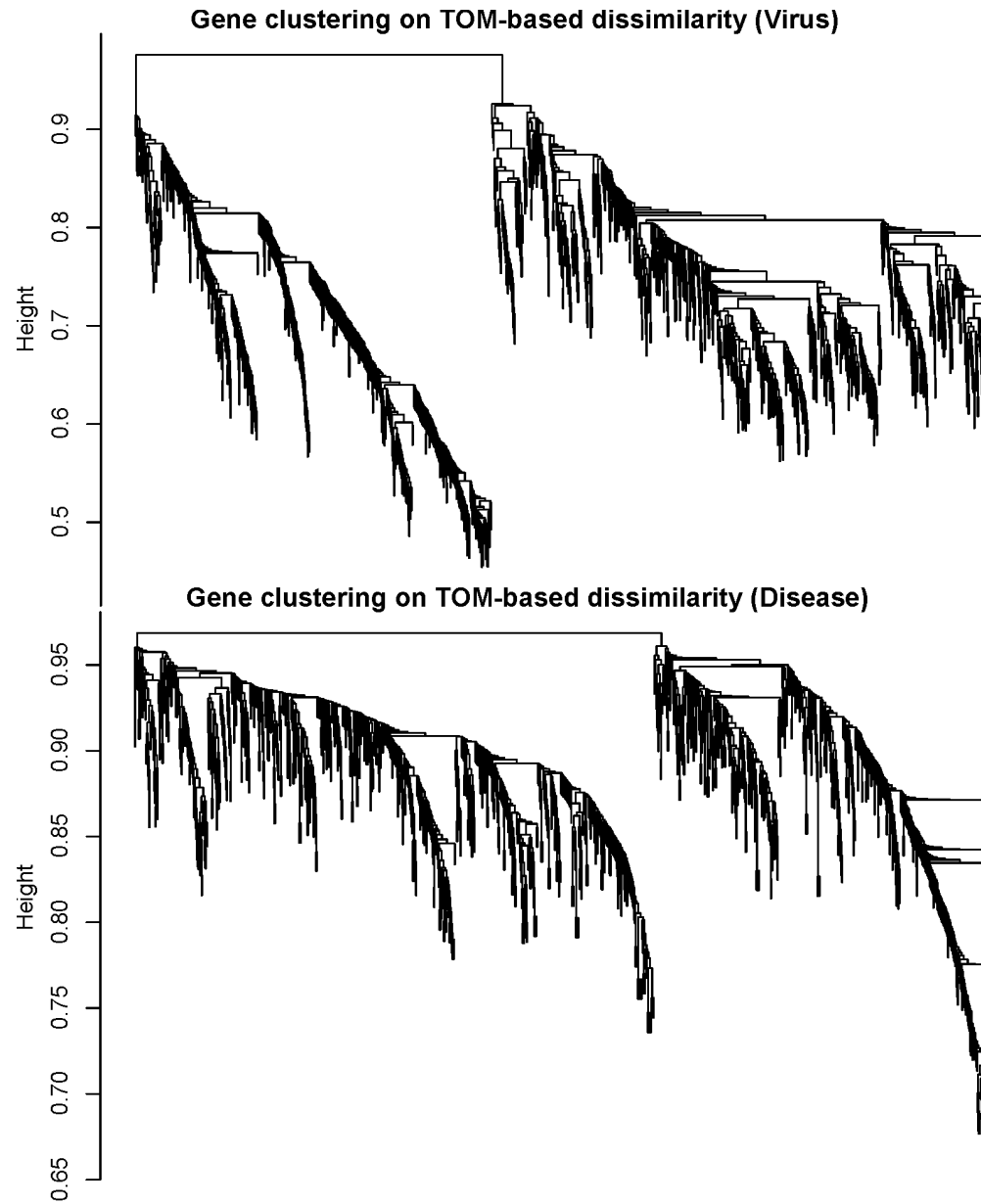


Hypothesis

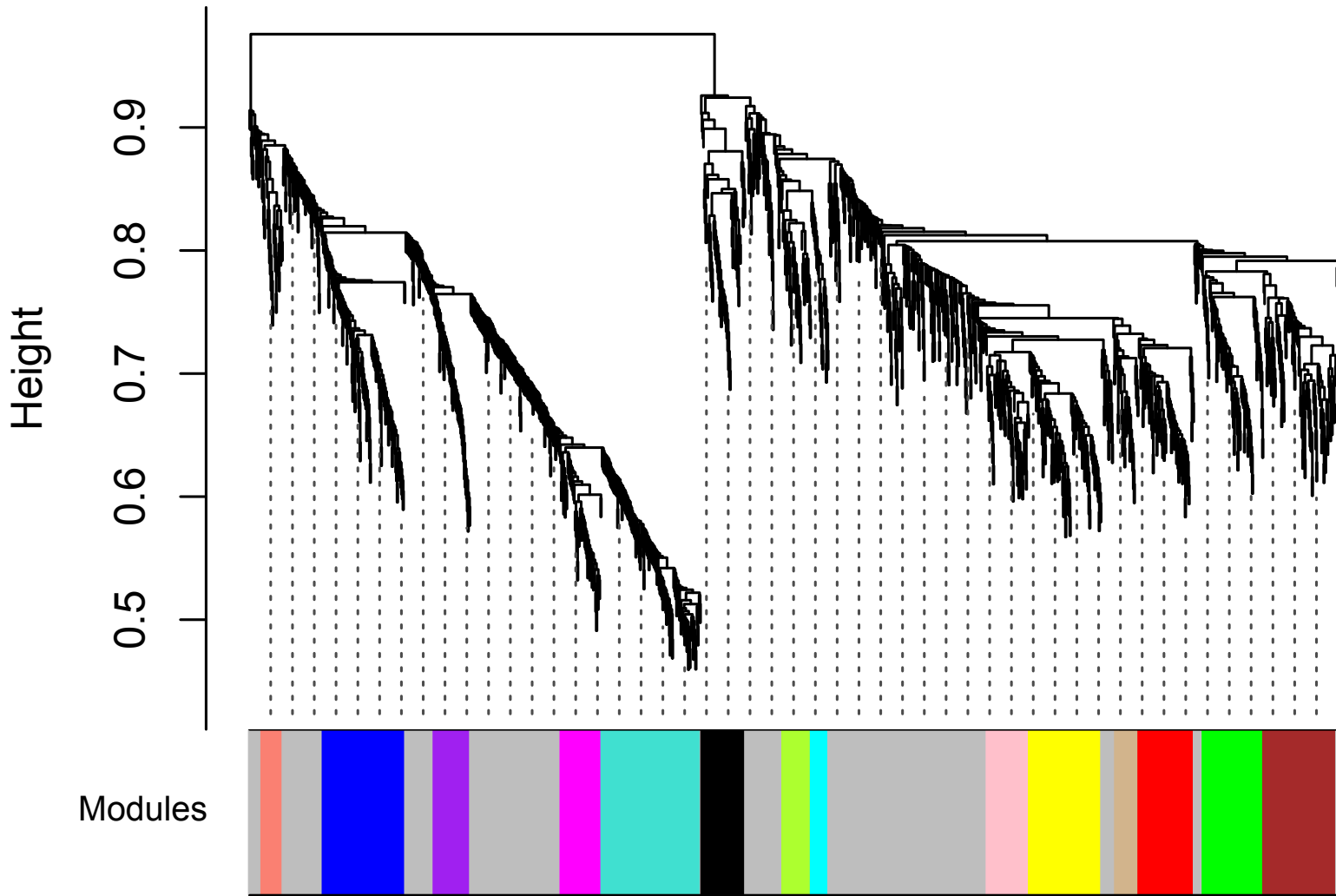
Common networks may represent vital evolutionarily conserved components of the coral stress response



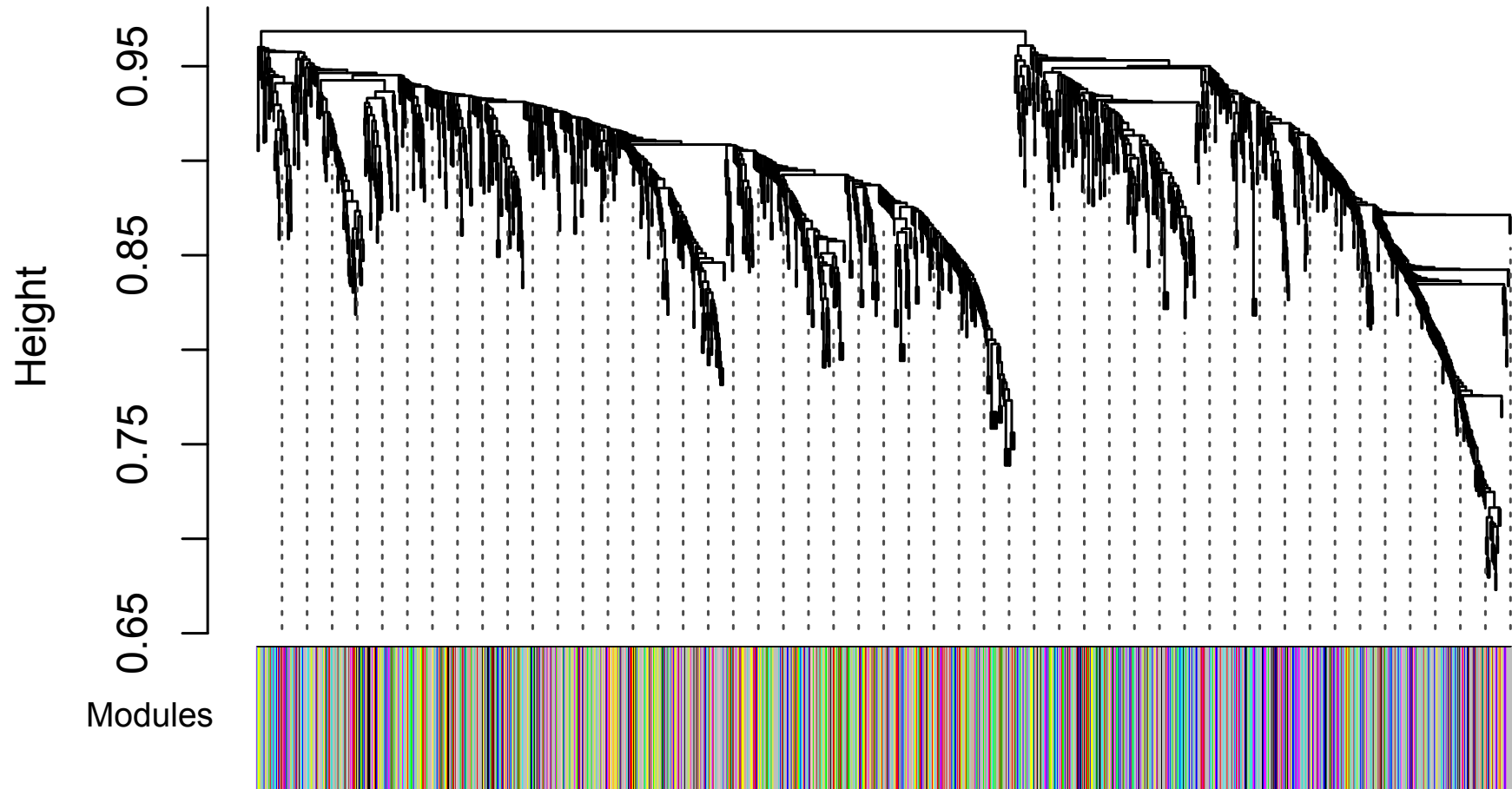
Identify gene networks in both data sets



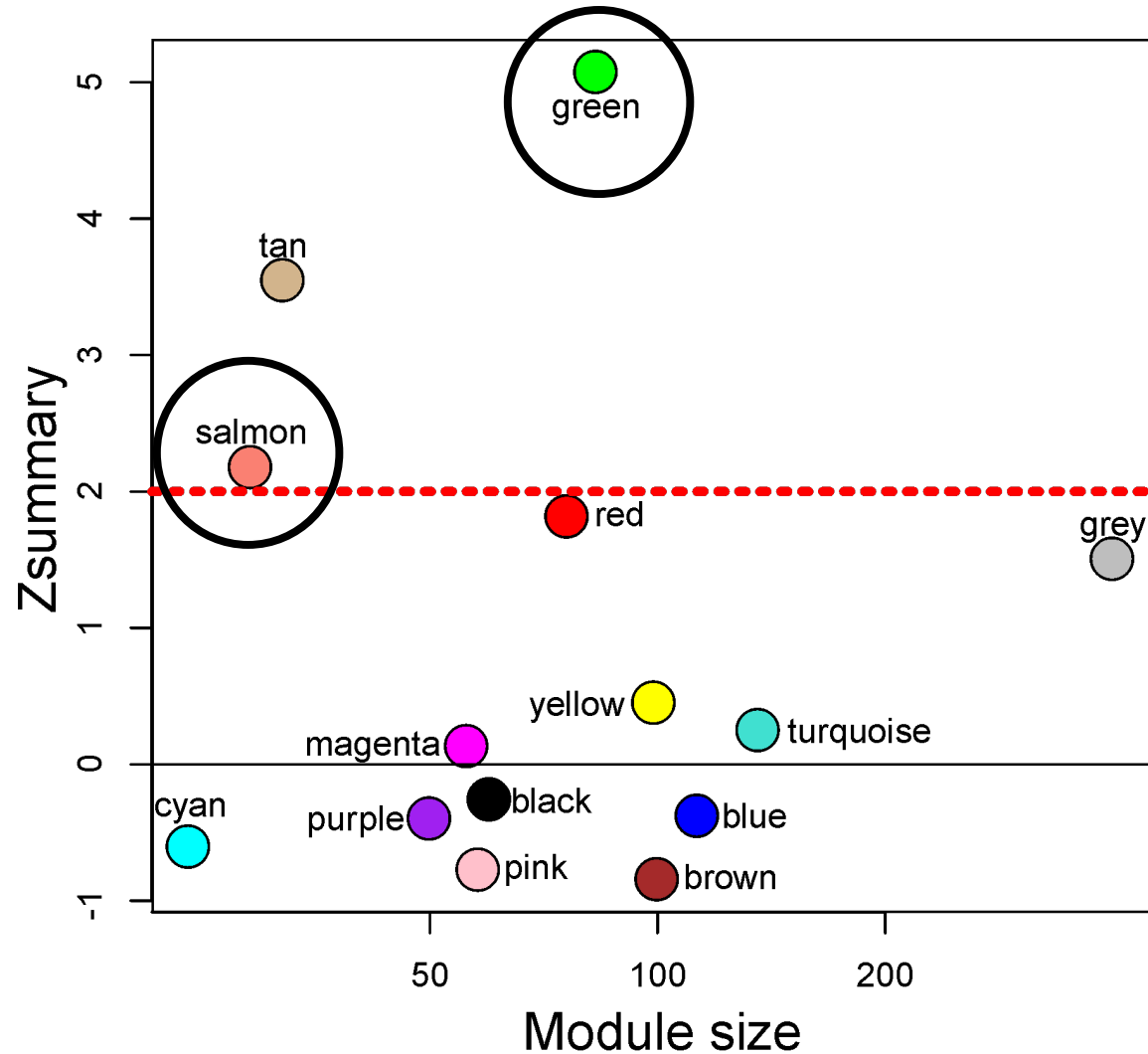
Assign modules to the virus data set



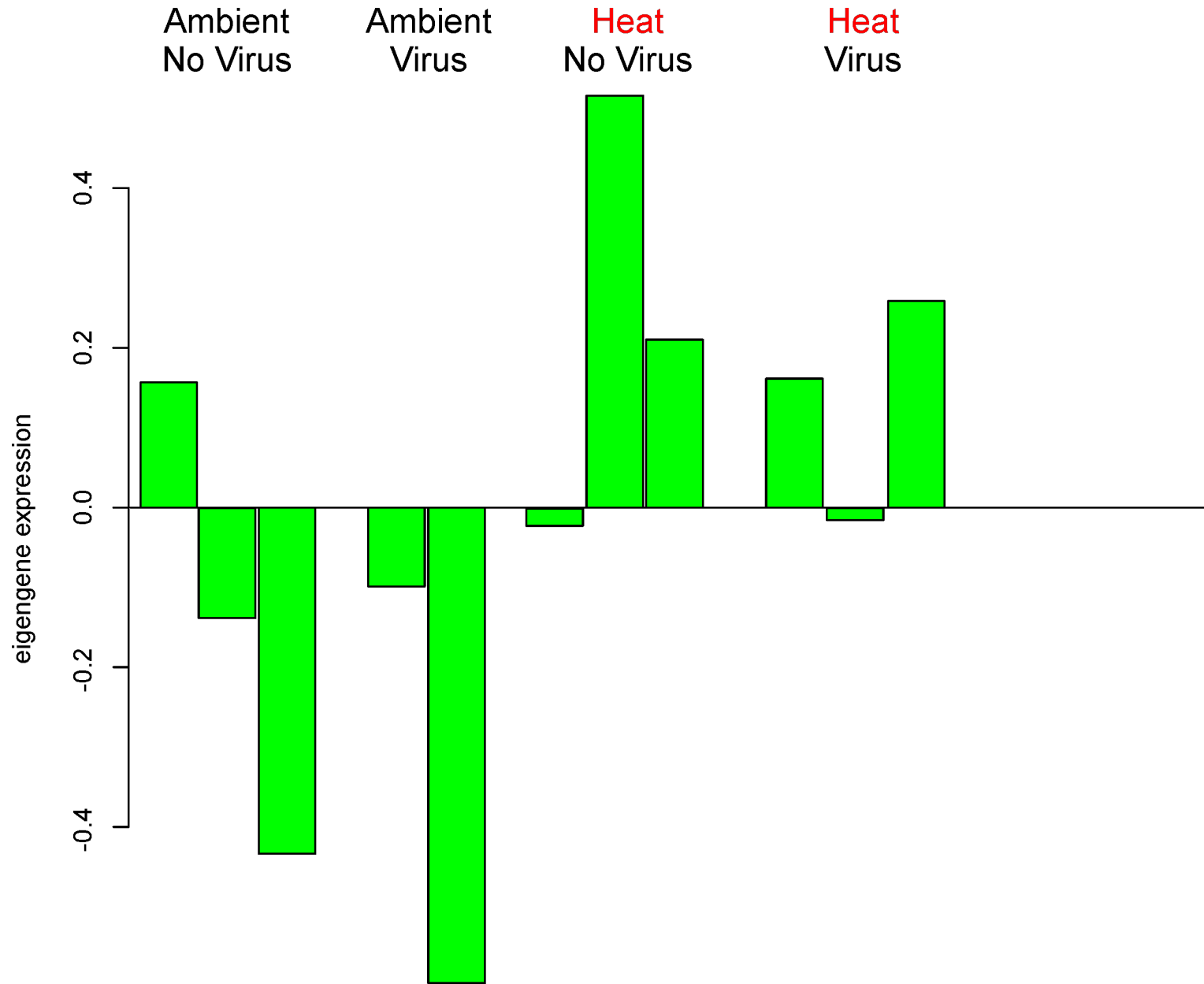
Impose the virus modules on the disease data set



Three modules were significantly preserved between the data sets



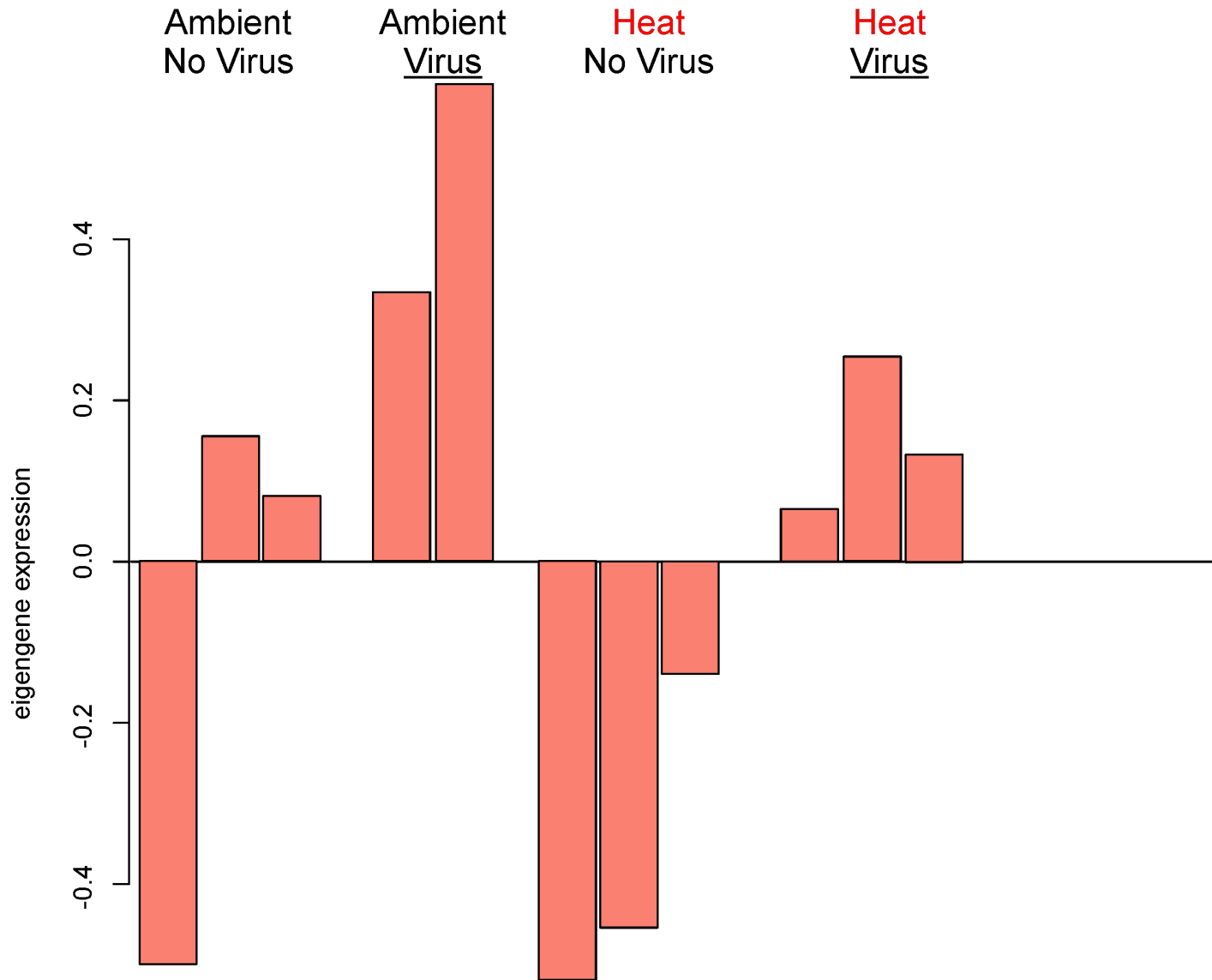
The green module correlates with heat stress



Functions include regulation of cell division, transcription, and protein modification

Annotation	Putative Function
voltage-gated sodium channel alpha, invertebrate	action potential regulation
ATP-binding cassette, subfamily A (ABC1), member 2	iron homeostasis, urate metabolism
WW domain containing adaptor with coiled-coil	transcription regulation
Rho guanine nucleotide exchange factor 2	cell cycle regulation
transitional endoplasmic reticulum ATPase	DNA damage response, protein modification
polycystin 1	cell projection regulation
nuclear factor erythroid 2-related factor 1/3	transcription regulation
alpha-1,3(6)-mannosylglycoprotein beta-1,6-N-acetyl-glucosaminyltransferase	post-translational protein modification
ribosomal RNA-processing protein 1	rRNA processing
mannosyl-oligosaccharide glucosidase	post-translational protein modification

The salmon module correlates with viral challenge



Functions include members of the innate immune response

Annotation	Putative Function
purinergic receptor P2X, ligand-gated ion channel 7	inflammatory response
peptide deformylase	co-translational processing of bacterial proteins
large subunit ribosomal protein L11e	translation
sequestosome 1	apoptosis, autophagy
deoxyhypusine monooxygenase	post-translational protein modification (eIF-5A)
transglutaminase 1	extracellular peptide cross-linking
ubiquinol-cytochrome c reductase subunit 6	oxidative phosphorylation
cofilin	cytokinesis (response to virus)
U3 small nucleolar RNA-associated protein 16	rRNA processing
calbindin D28	cellular response to organic substance

