

# Illumina - all flavors

If this page isn't formatted well on your screen, try shrinking the left side bar.

[Here is a helpful document from Illumina discussing the compatibility between various library primers and various sequencing platforms and kits.](#)

## Canonical ILLUMINA library design as of June 2012 (all 5'-3'), "TruSeq V3": NOTE all sequences shown are TOP STRAND 5' to 3'

```
<P5 primer/capture site>
<IndexRead2>
<Read1 primer site>
<template - gDNA, RNA, amplicon, whatever>
<Read2 primer site>
<IndexRead1>
<P7 primer/capture site>
```

### If you'd like a different description, [this one from the Tufts core facility is quite good.](#)

NOTE THAT THE SHADED PORTIONS SHOULD NOT BE CHANGED if you are designing your own primers!! The only flexibility one has is in the "template" section and in the two "index read" sections. Every other nucleotide shown matters as-is.

#### Single index adaptor design on a standard Illumina HiSeq or MiSeq run

1. P5 PCR primer/flowcell capture site:

```
AATGATACGGCGACCACCGAGA
```

2. IndexRead2:

NONE - as in **do NOT** put an index here. If you want to add an index here, use one of the "Dual-index" designs below.

3. Read1 primer site:

Either the small RNA sequencing primer site: (NEB: TCTACACGTTCTACAGTTCTACAGTCCGACGATCA [Illumina lists this but it is UNPROVEN: CAGGTTCTACAGTTCTACAGTCCGACGATCA]) OR the standard TruSeq Read 1 primer site: TCTACACTCTTTCCCTACACGACGCTCTTCCGATCT. Which to choose? The TruSeq Read 1 primer site is complementary to the Read 2 primer site, so if you are designing amplicons do NOT use the TruSeq Read 1 primer site, use the small RNA sequencing primer site.

4. The insert to be sequenced

5. Read2 primer site:

Then the Index read primer site: AGATCGGAAGAGCACACGTCTGAACTCCAGTCAC (**NOTE:** the initial A is from the dA tailing of the insert and is not included in the index primer or adaptor sequences; note also the reverse-complement of this is the Read 2 sequencing primer, but the Read 2 sequencing primer includes the T corresponding to the dA insert tail so sequencing starts with the insert)

6. IndexRead1:

The index sequence (usually 6 bp) - see many examples below in the **Barcodes** section. Within a lane, image analysis works best with as much base diversity as possible.

7. P7 PCR primer/flowcell capture site:

```
ATCTCGTATGCCGTCTTCTGCTTG
```

Here is an example of a read-pair from an RNA-seq library generated from the NEB small RNA kit with an insert size of 62 nt:

```
Read 1 sequence (note adaptor sequence starting with "AGATCGGAA...")
```

```
AAGGGATCATAGACGGTATTTCTATGTAACGAACAGTCGGGCGAGTCTCAGTGGGAGTTTCAGATCGGAAGAGCACACGTCTGAACTCCAGNACCGATG
ACCGAGATCTACAGTTCTACAGTTCTACAGTCCGACGATCAGGGATCATAGACGGTATTTCTATGTAACGAACAGTCGGGCGAGTCTCAGTGGGAGTTTC
5' SR adaptor:GUUCAGAGUUCUACAGUCCGACGAUC
```

```
Read 2 sequence, reverse complemented (note adaptor sequence RC ending with "...CCGACGATCA")
```

#### Dual-index TruSeq (NOT Nextera) adaptor design on a standard Illumina PE HiSeq or MiSeq run

1. P5 PCR primer/flowcell capture site:

```
AATGATACGGCGACCACCGAGATCTACAC
```

2. IndexRead2:

Optional. Example: TAGATCGC. This is called "IndexRead2" because it is read after index read 1. The GSAF does not normally sequence this barcode - please request if you need it read. We have little guidance to offer on designs other than to re-use the same sequences as in the Index Read 1 site - base diversity is your friend.

3. Read1 primer site:

The standard TruSeq Read 1 primer site: ACACTCTTCCCTACACGAGCTCTCCGATCT. We are not sure at this point whether the small RNA primer site is compatible with dual-indexes or not.

4. The remaining template elements are identical to the Single-index adaptor design above.
5. Note that an artifact of this design is that a SINGLE index (I7 index) TS library will read the sequence "TCTTTC..." as the I5 index if it is run in dual-index mode.

#### Dual-index Nextera adaptor design (we believe these are compatible with Illumina V3 PE HiSeq or V2/V3 MiSeq run)

```

Nextera DNA Sample Preparation Kit (Illumina) 1,2
Nextera transposase sequences (FC-121-1031, FC-121-1030)
5 TCGTCGGCAGCGTCAGATGTGTATAAGAGACAG
      (a) Read 1 -->
5 GTCTCGTGGGCTCGGAGATGTGTATAAGAGACAG
      (d) Read 2 -->

Nextera Index Kit - PCR primers (FC-121-1012, FC-121-1011)
5 AATGATACGGCGACCCAGAGATCTACAC[i5]TCGTCGGCAGCGTC
      (c) i5 Index read -->
5 CAAGCAGAAGACGGCATACGAGAT[i7]GTCTCGTGGGCTCGG
      <-- i7 Index read (b)

```

Nextera codes for entry on sample sheet:

i5 bases in adapter	Nextera DNA i5 index name	Nextera XT i5 index name	Nextera Enrichment i5 index name	HiSeq 2500 and MiSeq i5 bases for entry on sample sheet	NextSeq and HiSeq 4000 i5 bases for entry on sample sheet
TAGATCGC	N501	S501	E501	TAGATCGC	GCGATCTA
CTCTCTAT	N502	S502	E502	CTCTCTAT	ATAGAGAG
TATCCTCT	N503	S503	E503	TATCCTCT	AGAGGATA
AGAGTAGA	N504	S504	E504	AGAGTAGA	TCTACTCT
GTAAGGAG	N505	S505	E505	GTAAGGAG	CTCCTTAC
ACTGCATA	N506	S506	E506	ACTGCATA	TATGCAGT
AAGGAGTA	N507	S507	E507	AAGGAGTA	TACTCCTT
CTAAGCCT	N508	S508	E508	CTAAGCCT	AGGCTTAG
i7 bases in adapter	Nextera DNA i7 index name	Nextera XT i7 index name	Nextera Enrichment i7 index name	i7 bases for entry on sample sheet (HiSeq, MiSeq, or NextSeq)	
TCGCCTTA	N701	N701	N701	TAAGGCCA	
CTAGTACG	N702	N702	N702	CGTACTAG	
TTCTGCCT	N703	N703	N703	AGGCAGAA	
GCTCAGGA	N704	N704	N704	TCCTGAGC	
AGGAGTCC	N705	N705	N705	GGACTCCT	
CATGCCTA	N706	N706	N706	TAGGCATG	
GTAGAGAG	N707	N707	N707	CTCTCTAC	
CCTCTCTG	N708	N708	N708	CAGAGAGG	
AGCGTAGC	N709	N709	N709	GCTACGCT	
CAGCCTCG	N710	N710	N710	CGAGGCTG	
TGCCTCTT	N711	N711	N711	AAGAGGCA	
TCCTCTAC	N712	N712	N712	GTAGAGGA	

### Barcodes (also known as Indexes)

NOTE: Illumina barcodes (indexes) have varied significantly over time NOT ONLY in their sequence but also in WHERE they are placed in the sequencing construct.

The GSAF expects indexes to be in the 3' end of the final sequencing construct, between the Index read sequencing primer site and the P7 PCR primer site. If you are using dual-indexed samples with an additional barcode between the P5 bridge PCR primer site and the Read 1 sequencing primer site, we can easily accommodate that on a run but do not normally do so - you need to tell us.

The GSAF uses the following names for the following barcodes. Note that these sequences are shown 5'-3' when the P5 sequence is on the left. In other words, here is the first barcode shown in the context of the full 3'-end adaptor construct:

GATCGGAAGAGCACACGTCTGAACTCCAGTCACATCACGATCTCGTATGCCGTCTTCTGCTTG

Sequence	TruSeq name	NEXTFlex number
ATCACG	TSBC01	NFBC07
CGATGT	TSBC02	NFBC01
TTAGGC	TSBC03	NFBC08
TGACCA	TSBC04	NFBC02
ACAGTG	TSBC05	NFBC03
GCCAAT	TSBC06	NFBC04
CAGATC	TSBC07	NFBC05
ACTTGA	TSBC08	NFBC09
GATCAG	TSBC09	NFBC10
TAGCTT	TSBC10	NFBC11
GGCTAC	TSBC11	NFBC12
CTTGTA	TSBC12	NFBC06
AGTCAA	TSBC13	NFBC13
AGTTCC	TSBC14	NFBC14
ATGTCA	TSBC15	NFBC15
CCGTCC	TSBC16	NFBC16
GTAGAG	TSBC17	NFBC17
GTCCGC	TSBC18	NFBC18
GTGAAA	TSBC19	NFBC19
GTGGCC	TSBC20	NFBC20
GTTTCG	TSBC21	NFBC21
CGTACG	TSBC22	NFBC22
GAGTGG	TSBC23	NFBC23
GGTAGC	TSBC24	NFBC24
ACTGAT	TSBC25	NFBC25
ATGAGC	TSBC26	NFBC26
ATTCTT	TSBC27	NFBC27
CAAAAG	TSBC28	NFBC28
CAACTA	TSBC29	NFBC29
CACCGG	TSBC30	NFBC30
CACGAT	TSBC31	NFBC31
CACTCA	TSBC32	NFBC32
CAGGCG	TSBC33	NFBC33
CATGGC	TSBC34	NFBC34
CATTTT	TSBC35	NFBC35

CCAACA	TSBC36	NFBC36
CGGAAT	TSBC37	NFBC37
CTAGCT	TSBC38	NFBC38
CTATAC	TSBC39	NFBC39
CTCAGA	TSBC40	NFBC40
GTGATC	NEB40	
GACGAC	TSBC41	N/A
GCGCTA	N/A	NFBC41
TAATCG	TSBC42	NFBC42
TACAGC	TSBC43	NFBC43
TATAAT	TSBC44	NFBC44
TCATTC	TSBC45	NFBC45
TCCCGA	TSBC46	NFBC46
TCGAAG	TSBC47	NFBC47
TCGGCA	TSBC48	NFBC48

NOTE that TSBC41 is hamming distance 2 away from both TSBC31 and TSBC11; all others are hamming distance  $\geq 3$ .

Some additional 5 bp barcodes can be found here: <http://comailab.genomecenter.ucdavis.edu/index.php/Barcodes>

After exhaustive searching of all 4096 6-mers, the following table is all remaining 6 bp barcodes that have hamming distance of at least 3 from each other and the table above of 49 barcodes (NOTE: these have NOT been tested on the sequencer as of 2/7/12):

Sequence	GSAF name	
AAACAC	UTBC50	
TGAAGG	UTBC51	
AACATA	UTBC52	
CGCGTC	UTBC53	
GATACA	UTBC54	
GGTGTG	UTBC55	
TAAGAA	UTBC56	
AGCGAG	UTBC57	
CGGTTA	UTBC58	
AGCTTT	UTBC59	
TGGTCT	UTBC60	
TATCCC	UTBC61	
TGTCGT	UTBC62	
CCCCAC	UTBC63	
ATACGA	UTBC64	
CCCTTG	UTBC65	
ACCGGC	UTBC66	
TFACTG	UTBC67	
GGAACT	UTBC68	
GTTATT	UTBC69	

AAAAGT	UTBC70	
AAGGGA	UTBC71	
AAGTAT	UTBC72	
ACATCT	UTBC73	
ACGATT	UTBC74	
ACGCCG	UTBC75	
ACTCTC	UTBC76	
AGAATC	UTBC77	
ATTGGG	UTBC78	
CCGCGT	UTBC79	
CGCCCT	UTBC80	
CTGCAG	UTBC81	
GAAGTT	UTBC82	
GCACCC	UTBC83	
GCAGGA	UTBC84	
GCCGCG	UTBC85	
GGCGGT	UTBC86	
GTATTA	UTBC87	
TACGTG	UTBC88	
TCACAT	UTBC89	
TCTATA	UTBC90	
TGCAAA	UTBC91	
TGGCAC	UTBC92	
TGTTAG	UTBC93	
TTCTAT	UTBC94	
GGTACG	UTBC95	

**Excruciating details - USE WITH CAUTION - RNA PCR primers are NOT current as of Dec. 2011**

Oligonucleotide sequences for TruSeq™ RNA and DNA Sample Prep Kits1

TruSeq Universal Adapter

5 AATGATACGGCGACCACCGAGATCTACACTCTTTCCCTACACGACGCTCTTCCGATCT

TruSeq Adapters

barcode: ATCACG

TruSeq Adapter, Index 1

5 GATCGGAAGAGCACACGTCTGAACTCCAGTCACATCACGATCTCGTATGCCGTCTTCTGCTTG

TruSeq Adapter, Index 2

5 GATCGGAAGAGCACACGTCTGAACTCCAGTCACCGATGTATCTCGTATGCCGTCTTCTGCTTG

TruSeq Adapter, Index 3

5 GATCGGAAGAGCACACGTCTGAACTCCAGTCACTTAGGCATCTCGTATGCCGTCTTCTGCTTG

TruSeq Adapter, Index 4

5 GATCGGAAGAGCACACGTCTGAACTCCAGTCACTGACCAATCTCGTATGCCGTCTTCTGCTTG

TruSeq Adapter, Index 5

5 GATCGGAAGAGCACACGTCTGAACTCCAGTCACACAGTGATCTCGTATGCCGTCTTCTGCTTG

TruSeq Adapter, Index 6

5 GATCGGAAGAGCACACGTCTGAACTCCAGTCACGCAATATCTCGTATGCCGTCTTCTGCTTG

TruSeq Adapter, Index 7

5 GATCGGAAGAGCACACGTCTGAACTCCAGTCACCGATCATCTCGTATGCCGTCTTCTGCTTG

TruSeq Adapter, Index 8



RNA PCR Primer, Index 5 (RPI5)  
CAAGCAGAAGACGGCATAACGAGATCACTGTGTGACTGGAGTTCCTTGGCACCCGAGAATTCCA  
RNA PCR Primer, Index 6 (RPI6)  
CAAGCAGAAGACGGCATAACGAGATATGGCGTGACTGGAGTTCCTTGGCACCCGAGAATTCCA  
RNA PCR Primer, Index 7 (RPI7)  
CAAGCAGAAGACGGCATAACGAGATGATCTGGTGACTGGAGTTCCTTGGCACCCGAGAATTCCA  
RNA PCR Primer, Index 8 (RPI8)  
CAAGCAGAAGACGGCATAACGAGATCAAGTGTGACTGGAGTTCCTTGGCACCCGAGAATTCCA  
RNA PCR Primer, Index 9 (RPI9)  
CAAGCAGAAGACGGCATAACGAGATCTGATCGTGACTGGAGTTCCTTGGCACCCGAGAATTCCA  
RNA PCR Primer, Index 10 (RPI10)  
CAAGCAGAAGACGGCATAACGAGATAAGCTAGTGACTGGAGTTCCTTGGCACCCGAGAATTCCA  
RNA PCR Primer, Index 11 (RPI11)  
CAAGCAGAAGACGGCATAACGAGATGTAGCCGTGACTGGAGTTCCTTGGCACCCGAGAATTCCA  
RNA PCR Primer, Index 12 (RPI12)  
CAAGCAGAAGACGGCATAACGAGATTACAAGGTGACTGGAGTTCCTTGGCACCCGAGAATTCCA  
RNA PCR Primer, Index 13 (RPI13)  
CAAGCAGAAGACGGCATAACGAGATTTGACTGTGACTGGAGTTCCTTGGCACCCGAGAATTCCA  
RNA PCR Primer, Index 14 (RPI14)  
CAAGCAGAAGACGGCATAACGAGATGGAACGTGACTGGAGTTCCTTGGCACCCGAGAATTCCA  
RNA PCR Primer, Index 15 (RPI15)  
CAAGCAGAAGACGGCATAACGAGATTGACATGTGACTGGAGTTCCTTGGCACCCGAGAATTCCA  
RNA PCR Primer, Index 16 (RPI16)  
CAAGCAGAAGACGGCATAACGAGATGGACGGGTGACTGGAGTTCCTTGGCACCCGAGAATTCCA  
RNA PCR Primer, Index 17 (RPI17)  
CAAGCAGAAGACGGCATAACGAGATCTCTACGTGACTGGAGTTCCTTGGCACCCGAGAATTCCA  
RNA PCR Primer, Index 18 (RPI18)  
CAAGCAGAAGACGGCATAACGAGATGCGGAGCTGACTGGAGTTCCTTGGCACCCGAGAATTCCA  
RNA PCR Primer, Index 19 (RPI19)  
CAAGCAGAAGACGGCATAACGAGATTTTACCGTGACTGGAGTTCCTTGGCACCCGAGAATTCCA  
RNA PCR Primer, Index 20 (RPI20)  
CAAGCAGAAGACGGCATAACGAGATGGCCACGTGACTGGAGTTCCTTGGCACCCGAGAATTCCA  
RNA PCR Primer, Index 21 (RPI21)  
CAAGCAGAAGACGGCATAACGAGATCGAAACGTGACTGGAGTTCCTTGGCACCCGAGAATTCCA  
RNA PCR Primer, Index 22 (RPI22)  
CAAGCAGAAGACGGCATAACGAGATCGTACGGTGACTGGAGTTCCTTGGCACCCGAGAATTCCA  
RNA PCR Primer, Index 23 (RPI23)  
CAAGCAGAAGACGGCATAACGAGATCCACTCGTGACTGGAGTTCCTTGGCACCCGAGAATTCCA  
RNA PCR Primer, Index 24 (RPI24)  
CAAGCAGAAGACGGCATAACGAGATGCTACCGTGACTGGAGTTCCTTGGCACCCGAGAATTCCA  
RNA PCR Primer, Index 25 (RPI25)  
CAAGCAGAAGACGGCATAACGAGATATCAGTGTGACTGGAGTTCCTTGGCACCCGAGAATTCCA  
RNA PCR Primer, Index 26 (RPI26)  
CAAGCAGAAGACGGCATAACGAGATGCTCATGTGACTGGAGTTCCTTGGCACCCGAGAATTCCA  
RNA PCR Primer, Index 27 (RPI27)  
CAAGCAGAAGACGGCATAACGAGATAGGAATGTGACTGGAGTTCCTTGGCACCCGAGAATTCCA  
RNA PCR Primer, Index 28 (RPI28)  
CAAGCAGAAGACGGCATAACGAGATCTTTTGGTGACTGGAGTTCCTTGGCACCCGAGAATTCCA  
RNA PCR Primer, Index 29 (RPI29)  
CAAGCAGAAGACGGCATAACGAGATTAGTTGGTGACTGGAGTTCCTTGGCACCCGAGAATTCCA  
RNA PCR Primer, Index 30 (RPI30)  
CAAGCAGAAGACGGCATAACGAGATCCCGTGGTGACTGGAGTTCCTTGGCACCCGAGAATTCCA  
RNA PCR Primer, Index 31 (RPI31)  
CAAGCAGAAGACGGCATAACGAGATATCGTGGTGACTGGAGTTCCTTGGCACCCGAGAATTCCA  
RNA PCR Primer, Index 32 (RPI32)  
CAAGCAGAAGACGGCATAACGAGATTGAGTGGTGACTGGAGTTCCTTGGCACCCGAGAATTCCA  
RNA PCR Primer, Index 33 (RPI33)  
CAAGCAGAAGACGGCATAACGAGATCGCCTGGTGACTGGAGTTCCTTGGCACCCGAGAATTCCA  
RNA PCR Primer, Index 34 (RPI34)  
CAAGCAGAAGACGGCATAACGAGATGCCATGGTGACTGGAGTTCCTTGGCACCCGAGAATTCCA  
RNA PCR Primer, Index 35 (RPI35)  
CAAGCAGAAGACGGCATAACGAGATAAAATGGTGACTGGAGTTCCTTGGCACCCGAGAATTCCA  
RNA PCR Primer, Index 36 (RPI36)  
CAAGCAGAAGACGGCATAACGAGATTGTTGGTGACTGGAGTTCCTTGGCACCCGAGAATTCCA  
RNA PCR Primer, Index 37 (RPI37)  
CAAGCAGAAGACGGCATAACGAGATATCCCGTGACTGGAGTTCCTTGGCACCCGAGAATTCCA  
RNA PCR Primer, Index 38 (RPI38)  
CAAGCAGAAGACGGCATAACGAGATAGCTAGGTGACTGGAGTTCCTTGGCACCCGAGAATTCCA  
RNA PCR Primer, Index 39 (RPI39)  
CAAGCAGAAGACGGCATAACGAGATGTATAGGTGACTGGAGTTCCTTGGCACCCGAGAATTCCA  
RNA PCR Primer, Index 40 (RPI40)

CAAGCAGAAGACGGCATAACGAGATTCTGAGGTGACTGGAGTTCCTTGGCACCCGAGAATTCCA  
RNA PCR Primer, Index 41 (RPI41)  
CAAGCAGAAGACGGCATAACGAGATGTCGTGACTGGAGTTCCTTGGCACCCGAGAATTCCA  
RNA PCR Primer, Index 42 (RPI42)  
CAAGCAGAAGACGGCATAACGAGATCGATTAGTGACTGGAGTTCCTTGGCACCCGAGAATTCCA  
RNA PCR Primer, Index 43 (RPI43)  
CAAGCAGAAGACGGCATAACGAGATGCTGTAGTGACTGGAGTTCCTTGGCACCCGAGAATTCCA  
RNA PCR Primer, Index 44 (RPI44)  
CAAGCAGAAGACGGCATAACGAGATATTATAGTGACTGGAGTTCCTTGGCACCCGAGAATTCCA  
RNA PCR Primer, Index 45 (RPI45)  
CAAGCAGAAGACGGCATAACGAGATGAATGAGTGACTGGAGTTCCTTGGCACCCGAGAATTCCA  
RNA PCR Primer, Index 46 (RPI46)  
CAAGCAGAAGACGGCATAACGAGATTCGGGAGTGACTGGAGTTCCTTGGCACCCGAGAATTCCA  
RNA PCR Primer, Index 47 (RPI47)  
CAAGCAGAAGACGGCATAACGAGATCTCGAGTGACTGGAGTTCCTTGGCACCCGAGAATTCCA  
RNA PCR Primer, Index 48 (RPI48)  
CAAGCAGAAGACGGCATAACGAGATTCGGGAGTGACTGGAGTTCCTTGGCACCCGAGAATTCCA

#### Oligonucleotide sequences for Genomic DNA

##### Adapters

5' P-GATCGGAAGAGCTCGTATGCCGTCTTCTGCTTG  
5' AACTCTTTCCCTACACGACGCTCTTCCGATCT  
PCR Primers  
5' AATGATACGGCGACCACCGAGATCTACACTCTTTCCCTACACGACGCTCTTCCGATCT  
5' CAAGCAGAAGACGGCATAACGAGCTCTTCCGATCT

##### Genomic DNA Sequencing Primer

5' AACTCTTTCCCTACACGACGCTCTTCCGATCT

#### Paired End DNA oligonucleotide sequences

##### PE Adapters

5' P-GATCGGAAGAGCGGTTCAGCAGGAATGCCGAG  
5' AACTCTTTCCCTACACGACGCTCTTCCGATCT  
PE PCR Primer 1.0  
5' AATGATACGGCGACCACCGAGATCTACACTCTTTCCCTACACGACGCTCTTCCGATCT  
PE PCR Primer 2.0  
5' CAAGCAGAAGACGGCATAACGAGATCGGTCTCGGCATTCCTGCTGAACCGCTCTTCCGATCT

##### PE Read 1 Sequencing Primer

5' AACTCTTTCCCTACACGACGCTCTTCCGATCT

##### PE Read 2 Sequencing Primer

5' CGGTCTCGGCATTCCTGCTGAACCGCTCTTCCGATCT

#### Oligonucleotide sequences for the Multiplexing Sample Prep Oligo Only Kit

##### Multiplexing Adapters

5' P-GATCGGAAGAGCACACGTCT  
5' AACTCTTTCCCTACACGACGCTCTTCCGATCT  
Multiplexing PCR Primer 1.0  
5' AATGATACGGCGACCACCGAGATCTACACTCTTTCCCTACACGACGCTCTTCCGATCT  
Multiplexing PCR Primer 2.0

##### Multiplexing Read 1 Sequencing Primer

5' AACTCTTTCCCTACACGACGCTCTTCCGATCT

##### Multiplexing Index Read Sequencing Primer

5' GATCGGAAGAGCACACGTCTGAACTCCAGTCAC

##### Multiplexing Read 2 Sequencing Primer

5' GTGACTGGAGTTCAGACGTGTGCTCTTCCGATCT

##### PCR Primer, Index 1

CAAGCAGAAGACGGCATAACGAGATCGTGATGTGACTGGAGTTC

##### PCR Primer, Index 2

CAAGCAGAAGACGGCATAACGAGATACATCGGTGACTGGAGTTC

##### PCR Primer, Index 3

CAAGCAGAAGACGGCATAACGAGATGCCTAAGTGACTGGAGTTC

##### PCR Primer, Index 4



CAAGCAGAAGACGGCATAACGAGATTGGTCAGTGACTGGAGTTC  
 PCR Primer, Index 5  
 CAAGCAGAAGACGGCATAACGAGATCACTGTGTGACTGGAGTTC  
 PCR Primer, Index 6  
 CAAGCAGAAGACGGCATAACGAGATATGGCGTGACTGGAGTTC  
 PCR Primer, Index 7  
 CAAGCAGAAGACGGCATAACGAGATGATCTGGTGACTGGAGTTC  
 PCR Primer, Index 8  
 CAAGCAGAAGACGGCATAACGAGATTCAAGTGTGACTGGAGTTC  
 PCR Primer, Index 9  
 CAAGCAGAAGACGGCATAACGAGATCTGATCGTGACTGGAGTTC  
 PCR Primer, Index 10  
 CAAGCAGAAGACGGCATAACGAGATAAGCTAGTGACTGGAGTTC  
 PCR Primer, Index 11  
 CAAGCAGAAGACGGCATAACGAGATGTAGCCGTGACTGGAGTTC  
 PCR Primer, Index 12  
 CAAGCAGAAGACGGCATAACGAGATTACAAGTGACTGGAGTTC

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Nextera adaptor style:

```

5'-TCGTCGGCAGCGTCAGATGTGTATAAGAGACAGT-3'
      |||
3'-CAGAGCACCCGAGCCTCTACACATATTCTCTGTC-5'
  
```

TruSeq truncated adaptor:

```

5'-ACACTCTTTCCCTACACGACGCTCTTCCGATCT-3'
      |||
3'-CACTGACCTCAAGTCTGCACACGAGAAGGCTAGA-5'
  
```

Nextera adaptor style, with primers overlaid:  
 First cycle:

```

GGCTCGGGTGCTCTG CAAAGC TAGAGCATAACGGCAGAAGACGAAC-5'
      5'-TCGTCGGCAGCGTCAGATGTGTATAAGAGACAGT-3'   <UNKNOWN>   5'-
CTGTCTCTTTATACACATCTCCGAGCCACGAGAC-3'
      |||
      3'-CAGAGCACCCGAGCCTCTACACATATTCTCTGTC-5'   <UNKNOWN>   3'-
TGACAGAGAATATGTGTAGACTGCGACGGCTGCT-5'
  
```

Second and subsequent cycles:

```

AATGATACGGCGACCACCGAGATCTACAC ATCACG TCGTCGGCAGCGTC
      3'-AGCAGCCGTGCGAGTCTACACATATTCTCTGTCA   <UNKNOWN>   3'-
TGACAGAGAATATGTGTAGACTGCGACGGCTGCT-5'
  
```

## Cautions, common mistakes, and lessons learned from failure

1. Assembling the P7 side adaptor or primer wrong - the key thing to note is that the "canonical designs" are shown 5' to 3' across the entire finished sequencing construct. So if you're designing a reverse primer for the P7 side you have to use the reverse complement of ALL 3 DESIGN ELEMENTS (flow cell binding site, barcode, and sequencing primer site) and make sure they're in the right order.
2. Incorrect P5 dual-index design - the "ACAC" motif in the single index design MUST be repeated on both sides of an index within P5 - see the "dual index" designs specifically.
3. Reverse complement barcode sequences in either P5 or P7 side indexes, especially from amplicons - the fact that the Illumina sequencers read i5 differently is a pain - pay attention to that when submitting barcode sequences that will wind up in a sample sheet. And remember that the i7 index is read "forward, top strand" of the canonical design, which is reverse complement of the sequence that appears in a reverse primer used when creating a library.